

# A checklist of chromosome numbers and a review of karyotype variation in Odonata of the world

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## Abstract

The ancient insect order Odonata is divided into three suborders: Anisoptera and Zygoptera with approximately 3000 species worldwide each, and Anisozygoptera with only four extant species in the relict family Epiophlebiidae. An updated list of Odonata species studied regarding chromosome number, sex chromosome mechanism and the occurrence of m-chromosomes (= microchromosomes) is given. Karyotypes of 607 species (198 genera, 23 families), covering approximately 10% of described species, are reported: 423 species (125 genera, 8 families) of the Anisoptera, 184 species (72 genera, 14 families) of the Zygoptera, and one species of the Anisozygoptera. Among the Odonata, sex determination mechanisms in males can be of X(0), XY and  $X_1X_2Y$  types, and diploid chromosome numbers can vary from 6 to 41, with a clear mode at  $2n = 25$  (60%) and two more local modes at  $2n = 27$  (21%) and  $2n = 23$  (13%). The karyotype  $2n = 25(24A + X)$  is found in each of the three suborders and is the most typical (modal) in many families, including the best-covered Libellulidae, Corduliidae (Anisoptera), Lestidae, Calopterygidae, and Platycnemididae (Zygoptera). This chromosome set is considered ancestral for the Odonata in general. Chromosome rearrangements, among which fusions and fissions most likely



predominated, led to independent origins of similar karyotypes within different phylogenetic lineages of the order. The karyotype  $2n = 27(26A + X)$  prevails in Aeshnidae and Coenagrionidae, whereas the karyotype  $2n = 23(22A + X)$  is modal in Gomphidae and Chlorocyphidae, in both pairs of families one being from the Anisoptera while the other from the Zygoptera.

### Keywords

Chromosome numbers, damselflydragons, damselflies, dragonflies, m-chromosomes, sex chromosome mechanisms

## Introduction

The order Odonata, which comprises slightly more than 6,000 described species worldwide, is one of the most ancient among winged insects (Pterygota), dating from the Permian (Grimaldi and Engel 2005). Extant Odonata include two main suborders with approximately 3,000 species each, the Zygoptera or damselflies with about 308 genera and the Anisoptera or true dragonflies with about 344 genera. Within these suborders, up to 21 and 11 families (and sometimes more), respectively, are currently recognized. The third suborder, the Anisozygoptera or damselflydragons, includes only one genus *Epiophlebia* Calvert, 1903 with four extant species in the relict family Epiophlebiidae. A substantial body of evidence indicates that Anisoptera and Zygoptera are each monophyletic, and Zygoptera are sister to *Epiophlebia* plus Anisoptera (Rehn 2003; Kalkman et al. 2008; Dijkstra et al. 2013, 2014; Schorr and Paulson 2020).

The field of Odonata cytogenetics was heavily influenced by Bastiaan Kiauta, who has published dozens of papers and analyzed karyotypes of about 260 species and subspecies of this group (see References and Table 1). During the years that have passed since the publication of chromosome number checklist of Odonata (Kiauta 1972c), approximately 90 chromosome papers have been published. The number of examined species has since increased by more than 2.3 times, and now it seems appropriate to publish an updated list. In this review article, all data available today are presented in two tables and one figure. Table 1 includes all species studied so far cytogenetically and compiles data on their chromosome numbers, sex chromosome mechanisms and the occurrence of the so-called m-chromosomes (= microchromosomes). Table 2 summarizes data presented in Table 1 and shows the family-level variability of the above-mentioned traits (except m-chromosomes, since data on their presence or absence in specific species are often questionable) together with the most characteristic (modal) karyotypes for each of the families explored. On the Fig. 1, the modal karyotypes are mapped onto phylogenetic tree of Odonata families taken from Bybee et al. (2016) who in turn redrawn and synthesized it from Dijkstra et al. (2014) and Carle et al. (2015). In the final section of the review, the main characteristics of Odonata karyotypes are briefly discussed and prospects for future research are outlined.



**Table 1.** Cytogenetically analyzed species of Odonata and their main karyotype characteristics (chromosome numbers, sex chromosomes, m-chromosomes).

Taxon	Karyotype formula 2n ♂	m-chromo- somes	Country	References
ANISOZYGOPTERA				
EPIOPHLEBIOIDEA				
<b>Epiophlebiidae</b>				
1. <i>Epiophlebia superstes</i> Selys, 1889	25(24A+X)	–	Japan	Oguma 1951
ANISOPTERA				
AESHNOIDEA				
<b>Aeshnidae</b>				
2. <i>Aeshna caerulea</i> (Ström, 1783)	24(22A+neo-XY)	–	Finland	Oksala 1943
3. <i>A. canadiensis</i> Walker, 1908	27(26A+X)	+	USA	Cruden 1968
4. <i>A. clepsydra</i> Say, 1839	27(26A+X)	+	USA	Hung 1971
5. <i>A. crenata</i> Hagen, 1856	27(26A+X)	+	Finland	Oksala 1939a, 1943, 1944, 1952
	– » –	–	Russia	Perepelov and Bugrov 2002
6. <i>A. cyanea</i> (Müller, 1764)	27(26A+X)	+	Finland	Oksala 1943
	– » –	+	Netherlands	Kiauta 1969a
7. <i>A. grandis</i> (Linnaeus, 1758)	27(26A+X)	+	Former USSR	Fuchsówna and Sawczyńska 1928
	25(24A+X)	+	Former USSR	Makalowskaja 1940
	26(24A+neo-XY)	+	Finland	Oksala 1939a, 1943, 1944, 1945
	– » –	+	Netherlands	Kiauta 1967a–d 1968a, b, 1969a
	– » –	+	Russia	Perepelov and Bugrov 2002
	25(24A+X)	–	Finland	Nokkala et al. 2002
8. <i>A. isocles</i> (Müller, 1767)	27(26A+X)	–	USA	Kiauta 1978 as <i>Anaciaeschna isosceles</i> (Müller, 1767)
	25(24A + X)	+	Russia	Kuznetsova et al. 2020b
9. <i>A. juncea</i> (Linnaeus, 1758)	26(24A+neo-XY)	+	Finland	Oksala 1939a, 1943, 1944
	– » –	+	Former USSR	Makalowskaja 1940
	27(26A+X)	+	Italy	Kiauta 1971a
	26(24A+neo-XY)	+	Russia	Perepelov and Bugrov 2002
10. <i>A. mixta</i> Latreille, 1805	27(26A+X)	+	Netherlands	Kiauta 1969a
	25(24A+X)	+	India	Sandhu and Malhotra 1994a
	– » –	+	India	Sharma and Durani 1995
	27(26A+X)	+	Russia	Perepelov and Bugrov 2001b
11. <i>A. nigroflava</i> Martin, 1909	27(26A+X)	+	Japan	Katatani 1987
	– » –	–	Russia	Perepelov and Bugrov 2002
12. <i>A. palmata</i> Hagen, 1856	27(26A+X)	+	USA	Cruden 1968
13. <i>A. serrata</i> Hagen, 1856	26(24A+neo-XY)	+	Finland	Oksala 1943 as <i>A. osiliensis</i> Mierzejewski, 1913 and <i>A. s. fennica</i> Valle, 1938
14. <i>A. subarctica</i> Walker, 1908	27(26A+X)	+	USA	Oksala 1939a, 1943, 1952 as <i>A. s. elisabethae</i> Djakonov, 1922
	– » –	+	Switzerland	Kiauta and Kiauta 1980a as <i>A. s. elisabethae</i>
15. <i>A. umbrosa</i> Walker, 1908	27(26A+X)	+	USA	Cruden 1968 as <i>A. u. occidentalis</i> Walker, 1908 and <i>A. u. umbrosa</i> Walker, 1908
16. <i>A. verticalis</i> Hagen, 1861	27(26A+X)	+	USA	Hung 1971
17. <i>A. viridis</i> Eversmann, 1836	26(24A+neo-XY)	+	Finland	Oksala 1943
	– » –	+	Russia	Perepelov et al. 1998
18. <i>A. walkeri</i> Kennedy, 1917	27(26A+X)	+	USA	Cruden 1968
19. <i>Anaciaeschna jaspidea</i> (Burmeister, 1839)	27(26A+X)	+	India	Walia and Sandhu 1999
20. <i>Anax amazili</i> (Burmeister, 1839)	27(26A+X)	–	Argentina	Capitulo et al. 1991
	– » –	+	Argentina	Mola et al. 1999
21. <i>A. concolor</i> Brauer, 1865	27(26A+X)	+	Surinam	Kiauta 1979a
22. <i>A. ephippiger</i> (Burmeister, 1839)	13(12A+X)	+	India	Seshachar and Bagga 1962 as <i>Hemianax ephippiger</i> (Burmeister, 1839)
	14(12A+neo-XY)	+	India	Kiauta 1969a as <i>H. ephippiger</i>

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
23.	<i>A. guttatus</i> (Burmeister, 1839)	15(14A+X)	+	Nepal	Kiauta and Kiauta 1982
24.	<i>A. immaculiformis</i> Rambur, 1842	27(26A+X)	+	India	Sangal and Tyagi 1982
		— » —	+	India	Walia et al. 2018
25.	<i>A. imperator</i> Leach, 1815	27(26A+X)	+	France	Kiauta 1965, 1969a
		— » —	—	Kenya	Wasschner 1985
		— » —	+	Russia	Perepelov and Bugrov 2002
26.	<i>A. junius</i> (Drury, 1773)	27(26A+X)	+	USA	McGill 1904, 1907
		— » —	+	USA	Lefevre and McGill 1908
		— » —	—	Japan	Kichijo 1942a
		— » —	+	USA	Cruden 1968
		— » —	—		
27.	<i>A. longipes</i> Hagen, 1861	27(26A+X)	+	USA	Cruden 1968
28.	<i>A. nigrofasciatus</i> Oguma, 1915	27(26A+X)	+	Nepal	Kiauta 1974, 1975 ( <i>A. n. nigrolineatus</i> Fraser, 1935)
		25(24A+X)	+	India	Sandhu and Malhotra 1994a ( <i>A. n. nigrolineatus</i> )
		27(26A+X)	+	India	Walia and Sandhu 1999 ( <i>A. n. nigrolineatus</i> )
		— » —	+	India	Walia et al. 2018 ( <i>A. n. nigrolineatus</i> )
29.	<i>A. papuensis</i> (Burmeister, 1839)	27(26A+X)	+	Australia	Kiauta 1968c, 1969a as <i>Hemianax papuensis</i> (Burmeister, 1839)
30.	<i>A. parthenope</i> (Selys, 1839)	27(26A+X)	+	Japan	Omura 1957 as <i>A. parthenope julius</i> Brauer, 1865
		— » —	+	India	Thomas and Prasad 1986
		— » —	+	China	Zhu and Wu 1986 as <i>A. p. julius</i>
		25(24A+X)	+	Japan	Suzuki and Saitoh 1990 as <i>A. p. julius</i>
		27(26A+X)	+	India	Sandhu and Malhotra 1994a
31.	<i>Andaeschna unicolor</i> (Martin, 1908)	27(26A+X)	+	Bolivia	Cumming 1964 as <i>Aeshna</i> cf. <i>unicolor</i> Martin, 1908
32.	<i>Austroaeschna anacantha</i> Tillyard, 1908	27(26A+X)	+	Australia	Kiauta 1968c as <i>Acanthaeschna anacantha</i> (Tillyard, 1908)
33.	<i>A. multipunctata</i> (Martin, 1901)	27(26A+X)	+	Australia	Kiauta 1968c as <i>Acanthaeschna multipunctata</i> (Martin, 1901)
34.	<i>Basiaeschna janata</i> (Say, 1939)	25(24A+X)	—	USA	Cruden 1968
35.	<i>Boyeria maclachlani</i> (Selys, 1883)	27(26A+X)	+	Japan	Omura 1957
36.	<i>B. vinosa</i> (Say, 1839)	27(26A+X)	—	USA	Cruden 1968
37.	<i>Caliaeschna microstigma</i> (Schneider, 1845)	16(14A+neo-XY)	+	Greece	Kiauta 1972a
38.	<i>Castoraeschna castor</i> (Brauer, 1865)	27(26A+X)	+	Brazil	Kiauta 1972b
39.	<i>Cephalaeschna orbifrons</i> Selys, 1883	25(24A+X)	+	Nepal	Kiauta 1975
40.	<i>Cephalaeschna</i> sp.	25(24A+X)	+	India	Sandhu and Malhotra 1994a
41.	<i>Coryphaeschna adnexa</i> (Hagen, 1961)	27(26A+X)	—	Bolivia	Cumming 1964
42.	<i>C. perrensi</i> (McLachlan, 1887)	25(24A+X)	—	Argentina	Capitulo et al. 1991
		27(26A+X)	+	Argentina	Mola et al. 1999
		— » —	+	Argentina	De Gennaro et al. 2008
43.	<i>C. viriditas</i> Calvert, 1952	23(22A+X)	+	Surinam	Kiauta 1979a
44.	<i>Gynacantha bayadera</i> Selys, 1891	25(24A+X)	+	India	Walia 2007 as <i>G. milliardi</i> Fraser, 1936
		27(26A+X)	+		
45.	<i>G. hyalina</i> Selys, 1882	28(26A+XX)*	+	India	Tyagi 1978a, b
46.	<i>G. interioris</i> Williamson, 1923	26(24A+neo-XY)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
47.	<i>G. japonica</i> Bartenev, 1909	27(26A+X)	+	Japan	Omura 1957
48.	<i>Gynacanthaeschna sikkima</i> (Karsch, 1891)	27(26A+X)	+	India	Walia et al. 2016
49.	<i>Oplonaeschna armata</i> (Hagen, 1861)	27(26A+X)	+	Mexico	Kiauta 1970a
50.	<i>Planaeschna milnei</i> (Selys, 1883)	27(26A+X)	+	Japan	Kiauta 1968c, 1969a
51.	<i>Remartinia luteipennis</i> (Burmeister, 1839)	25(24A+X)	+	Surinam	Kiauta 1979a as <i>Coryphaeschna l. luteipennis</i> Burmeister, 1839
		27(26A+X)	+	Brazil	Ferreira et al. 1979 as <i>C. l. luteipennis</i>
52.	<i>Rhionaeschna bonariensis</i> (Rambur, 1842)	26(24A+neo-XY)	+	Argentina, Uruguay	Mola and Papeschi 1994 as <i>Aeschna bonariensis</i> Rambur, 1842



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
52.	<i>Rhionaeschna bonariensis</i> (Rambur, 1842)	— » —	+	Argentina, Uruguay	Mola 1995 as <i>A. bonariensis</i>
53.	<i>Rh. californica</i> (Calvert, 1895)	27(26A+X)	+	Canada	Kiauta 1973a as <i>Aeshna californica</i> Calvert, 1895
54.	<i>Rh. confusa</i> (Rambur, 1842)	27(26A+X)	+	Argentina, Uruguay	Mola and Papeschi 1994 as <i>Aeshna confuse</i> Rambur, 1842
		— » —	+	Argentina, Uruguay	Mola 1995 as <i>A. confuse</i>
55.	<i>Rh. diffinis</i> (Rambur, 1842)	21(20A+X)	+	Bolivia	Cumming 1964 as <i>Aeshna d. diffinis</i> Rambur, 1842
56.	<i>Rh. intricata</i> (Martin, 1908)	19(18A+X)	+	Bolivia	Cumming 1964 as <i>Aeshna intricata</i> Martin, 1908
57.	<i>Rh. peralta</i> (Ris, 1918)	27(26A+X)	+	Bolivia	Cumming 1964 as <i>Aeshna peralta</i> Ris, 1918
58.	<i>Rh. planaltica</i> (Calvert, 1845)	16(14A+neo-XY)	+	Argentina	Mola and Papeschi 1994 as <i>Aeschna</i> <i>cornigera planaltica</i> Calvert, 1952
		— » —	+	Argentina	Mola 1995 as <i>A. c. planaltica</i>
59.	<i>Staurophlebia reticulata</i> (Burmeister, 1839)	27(26A+X)	+	Brazil	Souza Bueno 1982 ( <i>S. r. reticulata</i> (Burmeister, 1839))
PETALUROIDEA					
Petaluridae					
60.	<i>Tachopteryx thoreyi</i> (Hagen, 1857)	19(18A+X)	+	USA	Cumming 1964
61.	<i>Tanypteryx hageni</i> (Selys, 1879)	17(16A+X)	+	USA	Cruden 1968
62.	<i>T. pryeri</i> (Selys, 1889)	17(16A+X)	+	Japan	Kichijo 1939, 1942a
63.	<i>Uropetala carovei</i> (White, 1846)	17(16A+X)**	+	New Zealand	Wolfe 1953
		25(24A+X)	+	New Zealand	Jensen and Mahanty 1978
		— » —	+	New Zealand	Jensen 1980
GOMPHOIDEA					
Gomphidae					
64.	<i>Anisogomphus bivittatus</i> (Selys, 1854)	23(22A+X)	+	India	Das 1956
		— » —	+	India	Walia and Chahal 2020
65.	<i>A. occipitalis</i> (Selys, 1854)	23(22A+X)	—	Nepal	Kiauta 1974, 1975
66.	<i>Aphylla edentata</i> Selys, 1869	23(22A+X)	—	Bolivia	Cumming 1964
67.	<i>A. producta</i> Selys, 1854	23(22A+X)	—	Bolivia	Cumming 1964
68.	<i>A. theodorina</i> (Navas, 1933)	23(22A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
69.	<i>A. williamsoni</i> (Gloyd, 1936)	23(22A+X)	+	USA	Kiauta and Brink 1978
70.	<i>Aphylla</i> sp.	23(22A+X)	+	Argentina	Mola 2007
71.	<i>Arigomphus lentulus</i> (Needham, 1902)	23(22A+X)	—	USA	Cruden 1968 as <i>Gomphus lentulus</i> Needham, 1902
72.	<i>A. pallidus</i> (Rambur, 1842)	23(22A+X)	—	USA	Cumming 1964 as <i>Gomphus pallidus</i> Rambur, 1842
73.	<i>A. submedianus</i> (Williamson, 1914)	23(22A+X)	—	USA	Cruden 1968 as <i>Gomphus</i> <i>submedianus</i> Williamson, 1914
74.	<i>Asiagomphus melaenops</i> (Selys, 1854)	23(22A+X)	+	Japan	Toyoshima and Hirai 1953 as <i>Gomphus melaenops</i> Selys, 1854
		— » —	+	Japan	Hirai 1956 as <i>G. melaenops</i>
		— » —	+	USA	Cruden 1968 as <i>G. melaenops</i>
75.	<i>Burmagomphus pyramidalis</i> Laidlaw, 1922	23(22A+X)	+	India	Tyagi 1977
76.	<i>Davidius nanus</i> (Selys, 1869)	23(22A+X)	—	Japan	Kichijo 1939, 1942a
77.	<i>Dromogomphus spinosus</i> (Selys, 1854)	23(22A+X)	+	USA	Cruden 1968
78.	<i>D. spoliatus</i> (Hagen, 1857)	23(22A+X)	+	USA	Cruden 1968
79.	<i>Epigomphus llama</i> Calvert, 1903	23(22A+X)	—	Bolivia	Cumming 1964
80.	<i>Erpetogomphus designatus</i> Hagen, 1857	23(22A+X)	+	USA	Cumming 1964
81.	<i>E. diadophis</i> Calvert, 1905	23(22A+X)	—	USA	Cumming 1964
82.	<i>E. ophibolus</i> Calvert, 1905	23(22A+X)	+	Mexico	Kiauta 1970a
83.	<i>Gomphoides</i> sp.	23(22A+X)	—	Bolivia	Cumming 1964
84.	<i>Gomphus confraternus</i> Selys, 1873	23(22A+X)	+	USA	Cruden 1968
85.	<i>G. exilis</i> Selys, 1854	23(22A+X)	+	USA	Cruden 1968
		— » —	+	Canada	Kiauta 1969a
86.	<i>G. graslini</i> Rambur, 1842	12(10A+neo-neo-XY)	+	France	Kiauta 1968d, 1969a



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
87.	<i>G. pulchellus</i> Selys, 1840	23(22A+X)	+	France	Kiauta 1973b
88.	<i>G. vulgatissimus</i> (Linnaeus, 1758)	23(22A+X)	–	Russia	Perepelov et al. 2001
89.	<i>Ictinogomphus rapax</i> (Rambur, 1942)	23(22A+X)	+	India	Asana and Makino 1935
		– » –	+	India	Makino 1935
		– » –	+	India	Kichijo 1942a
		– » –	+	India	Omura 1949, 1952, 1953
		– » –	+	India	Dasgupta 1957
90.	<i>Nepogomphus modestus</i> (Selys, 1878)	23(22A+X)	–	India	Walia et al. 2006
		– » –	–	India	Walia and Chahal 2014
91.	<i>Nihonogomphus ruptus</i> (Selys, 1858)	23(22A+X)	–	Russia	Perepelov et al. 2001
92.	<i>N. viridis</i> Oguma, 1926	23(22A+X)	+	Japan	Omura 1957
93.	<i>Nychogomphus duaricus</i> (Fraser, 1924)	22(20A+neo-XY)	+	India	Tyagi 1977
94.	<i>Octogomphus specularis</i> (Hagen, 1859)	23(22A+X)	–	USA	Cruden 1968
95.	<i>Onychogomphus forcipatus</i> (Linnaeus, 1758)	25(24A+X)	–	Finland	Oksala 1945
		22(20A+neo-XY)	–	Austria	Kiauta 1969a
		25(24A+X)	–		
96.	<i>O. saundersii</i> Selys, 1854	22(20A+neo-XY)	+	India	Tyagi 1977 ( <i>O. s. duaricus</i> Fraser, 1924)
97.	<i>Ophiogomphus bison</i> Selys, 1873	23(22A+X)	–	USA	Cruden 1968
		25(24A+X)	–		
98.	<i>O. cecilia</i> (Fourcroy, 1785)	24(22A+XX)*	–	Finland	Oksala 1945
		23(22A+X)	–	Russia	Perepelov et al. 1998
		– » –	–	Russia	Perepelov and Bugrov 2001a
99.	<i>O. colubrinus</i> Selys, 1854	23(22A+X)	–	USA	Cruden 1968
100.	<i>O. obscurus</i> Bartenev, 1909	23(22A+X)	–	Russia	Perepelov and Bugrov 2001b
101.	<i>O. occidentalis</i> Hagen, 1882	23(22A+X)	–	USA	Cruden 1968
102.	<i>O. rupinsulensis</i> (Walsh, 1862)	23(22A+X)	–	USA	Cruden 1968
103.	<i>Phanogomphus lividus</i> (Selys, 1854)	23(22A+X)	+	USA	Cruden 1968 as <i>Gomphus lividus</i> Selys, 1854
104.	<i>Ph. militaris</i> (Hagen, 1858)	23(22A+X)	–	USA	Cruden 1968 as <i>Gomphus militaris</i> Hagen, 1858
105.	<i>Ph. spicatus</i> (Selys, 1854)	23(22A+X)	+	USA	Cruden 1968 as <i>Gomphus spicatus</i> Selys, 1854
106.	<i>Paragomphus lineatus</i> (Selys, 1850)	23(22A+X)	–	Nepal	Kiauta 1974, 1975
		– » –	–	India	Walia and Chahal 2014
107.	<i>P. capricornis</i> (Förster, 1914)	23(22A+X)	–	Thailand	Kiauta and Kiauta 1983
108.	<i>Phyllocycla propinqua</i> Belle, 1972	21(20A+X)	–	Argentina	De Gennaro 2004
109.	<i>Phyllocycla</i> sp.	23(22A+X)	–	Bolivia	Cumming 1964
110.	<i>Phyllocycla</i> sp. 1	23(22A+X)	+	Argentina	Mola 2007
111.	<i>Phyllocycla</i> sp. 2	23(22A+X)	–	Argentina	Mola 2007
112.	<i>Phyllogomphoides undulatus</i> (Needham, 1944)	23(22A+X)	+	Surinam	Kiauta 1979a
113.	<i>Progomphus borealis</i> McLachlan, 1873	23(22A+X)	–	USA	Cruden 1968
114.	<i>P. intricatus</i> (Hagen, 1857)	23(22A+X)	–	Bolivia	Cumming 1964
115.	<i>P. obscurus</i> (Rambur, 1842)	23(22A+X)	–	USA	Cruden 1968
116.	<i>P. phyllochromus</i> Ris, 1918	23(22A+X)	+	Bolivia	Cumming 1964
117.	<i>Scalmogomphus bistrigatus</i> (Hagen, 1854)	23(22A+X)	–	Nepal	Kiauta 1974, 1975 as <i>Onychogomphus bistrigatus</i> (Hagen, 1854)
118.	<i>Shaogomphus postocularis</i> (Selys, 1869)	23(22A+X)	+	Japan	Omura 1957 as <i>Gomphus postocularis</i> Selys, 1869
		– » –	–	Russia	Perepelov et al. 2001 as <i>Gomphus epophthalmus</i> Selys, 1872
119.	<i>Sieboldius albardae</i> Selys, 1886	23(22A+X)	+	Japan	Omura 1957
120.	<i>Stylogomphus suzukii</i> (Matsumura, 1926)	23(22A+X)	+	Japan	Oguma 1930
		– » –	+	Japan	Kichijo 1942a
121.	<i>Stylurus flavipes</i> (Charpentier, 1825)	23(22A+X)	+	Russia	Perepelov and Bugrov 2001b
122.	<i>S. plagiatus</i> (Selys, 1854)	23(22A+X)	+	USA	Cruden 1968 as <i>Gomphus plagiatus</i> Selys, 1854
123.	<i>S. scudderi</i> (Selys, 1873)	23(22A+X)	–	USA	Cruden 1968 as <i>Gomphus scudderi</i> Selys, 1873



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
124.	<i>S. townesi</i> Gloyd, 1936	22(20A+neo-XY)	–	USA	Kiauta and Brink 1978 as <i>Gomphus townesi</i> Gloyd, 1936
125.	<i>Temnogomphus bivittatus</i> (Selys, 1854)	23(22A+X)	+	Nepal	Kiauta 1975
126.	<i>Trigomphus citimus</i> (Needham, 1931)	21(20A+X)	+	Japan	Toyoshima and Hirai 1953 ( <i>T. c. tabei</i> Asahina, 1949)
		– » –	+	Japan	Hirai 1956 ( <i>T. c. tabei</i> )
127.	<i>T. interruptus</i> (Selys, 1854)	19(18A+X)	+	Japan	Oguma 1930
		– » –	+	Japan	Toyoshima and Hirai 1953
		– » –	+	Japan	Hirai 1956
		– » –	+	Japan	Omura 1957
128.	<i>T. melampus</i> (Selys, 1869)	21(20A+X)	–	Japan	Oguma 1930, 1942 as <i>T. unifasciatus</i> (Oguma 1926)
129.	<i>Zonophora callipus</i> Selys, 1869	23(22A+X)	+	Surinam	Kiauta 1979a
LIBELLULOIDEA					
Macromiidae					
130.	<i>Didymops transversa</i> (Say, 1839)	25(24A+X)	+	USA	Cruden 1968
131.	<i>Epophthalmia frontalis</i> (Selys, 1871)	25(24A+X)	+	India	Dasgupta 1957 ( <i>E. f. frontalis</i> (Selys, 1871))
132.	<i>Macromia daimoji</i> Okumura, 1949	25(24A+X)	–	Japan	Katatani 1987
133.	<i>M. amphigenia</i> Selys, 1871	25(24A+X)	–	Russia	Perepelov and Bugrov 2001b ( <i>M. a. fraenata</i> Martin, 1906)
134.	<i>M. magnifica</i> (McLachlan, 1874)	25(24A+X)	+	USA	Cruden 1968
		– » –	–		
135.	<i>M. moorei</i> Selys, 1874	25(24A+X)	+	Nepal	Kiauta 1977
		– » –	+	India	Walia and Chahal 2018
Corduliidae					
136.	<i>Cordulia aenea</i> (Linnaeus, 1758)	25(24A+X)	–	Finland	Oksala 1939a
		– » –	–	Former USSR	Makalowskaja 1940
		– » –	–	Netherlands	Kiauta 1968b, 1969a
		– » –	–	Russia	Perepelov et al. 1998
		– » –	–	Bulgaria	Grozeva and Marinov 2007
		– » –	–	Russia	Kuznetsova et al. 2018
137.	<i>C. shurtleffi</i> Scudder, 1866	25(24A+X)	+	USA	Cruden 1968
		– » –	+	Canada	Kiauta 1973a
138.	<i>Dorocordulia libera</i> (Selys, 1871)	11(10A+X)	–	USA	Cruden 1968
		13(12A+X)	–		
		14(12A+neo-XY)	–	USA	Kiauta 1969a
		13(12A+X)	–		
139.	<i>Epicordulia princeps</i> (Hagen, 1861)	25(24A+X)	+	USA	Hung 1971
140.	<i>Epitheca bimaculata</i> (Charpentier, 1825)	25(24A+X)	–	Russia	Perepelov 2003
		– » –	–	Russia	Kuznetsova et al. 2018
141.	<i>E. canis</i> McLachlan, 1886	25(24A+X)	+	USA	Cruden 1968
142.	<i>E. cynosura</i> (Say, 1839)	19(18A+X)	–	USA	Cruden 1968
		21(20A+X)	–		
143.	<i>E. petechialis</i> (Muttkowski, 1911)	21(20A+X)	–	USA	Cumming 1964 as <i>Tetragoneuria petechialis</i> Muttkowski, 1911
144.	<i>E. semiaquea</i> (Burmeister, 1839)	25(24A+X)	–	USA	Cruden 1968
145.	<i>E. spinigera</i> (Selys, 1871)	25(24A+X)	+	USA	Cruden 1968
		27(26A+X)	–	USA	Hung 1971 as <i>Tetragoneuria spinigera</i> (Selys, 1871)
146.	<i>Procordulia grayi</i> (Selys, 1871)	25(24A+X)	+	New Zealand	Jensen 1980
147.	<i>P. smithii</i> (White, 1846)	25(24A+X)	+	New Zealand	Jensen 1980
148.	<i>Rialla villosa</i> Rambur, 1842	25(24A+X)	+	Argentina	De Gennaro 2004
149.	<i>Somatochlora alpestris</i> (Selys, 1840)	25(24A+X)	–	Switzerland	Kiauta and Kiauta 1980a
		27(26A+X)	+		
150.	<i>S. arctica</i> (Zetterstedt, 1840)	25(24A+X)	+	Russia	Perepelov 2003
151.	<i>S. borisi</i> Marinov, 2001	20(18A+XY)	–	Bulgaria	Grozeva and Marinov 2007
152.	<i>S. flavomaculata</i> (Van der Linden, 1825)	25(24A+X)	–	Former USSR	Makalowskaja 1940
		– » –	–	Russia	Perepelov 2003
		– » –	+	Russia	Kuznetsova et al. 2020b

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
153.	<i>S. graeseri</i> Selys, 1887	25(24A+X)	—	Russia	Perepelov et al. 2001
154.	<i>S. meridionalis</i> Nielsen, 1935	25(24A+X)	—	Slovenia	Kiauta and Kiauta 1995
		— » —	—	Bulgaria	Grozeva and Marinov 2007
155.	<i>S. metallica</i> (Van der Linden, 1825)	26(24A+XX)*	—	Finland	Oksala 1945
		25(24A+X)	—	Finland	Nokkala et al. 2002
		— » —	—	Finland	Grozeva and Marinov 2007
		— » —	—	Russia	Perepelov and Bugrov 2001b
156.	<i>S. semicircularis</i> (Selys, 1871)	25(24A+X)	—	USA	Cruden 1968
157.	<i>S. uchidai</i> Fürster, 1909	25(24A+X)	+	Japan	Oguma 1915, 1930
		— » —	+	Japan	Kichijo 1942b
158.	<i>S. viridiaenea</i> (Uhler, 1858)	25(24A+X)	—	Japan	Oguma 1915, 1930
		— » —	—	Japan	Kichijo 1942b
Libellulidae					
159.	<i>Acisoma panorpoides</i> Rambur, 1842	25(24A+X)	+	Bangladesh, India	Dasgupta 1957 ( <i>A. p. panorpoides</i> Rambur, 1842)
		— » —	+	Nepal	Kiauta 1975 ( <i>A. p. panorpoides</i> )
		— » —	+	Thailand	Kiauta and Kiauta 1983 ( <i>A. p. panorpoides</i> )
		— » —	+	India	Tyagi 1982
160.	<i>Aethriamanta brevipennis</i> (Rambur, 1842)	25(24A+X)	+	India	Dasgupta 1957
161.	<i>Anatya guttata</i> (Erichson, 1848)	25(24A+X)	—	Surinam	Kiauta 1979a
162.	<i>Atoconeura biordinata</i> Karsch, 1899	21(20A+X)	+	Sudan	Wasscher 1985
163.	<i>Brachydiplax chalybea</i> Brauer, 1868	25(24A+X)	+	India	Dasgupta 1957
		— » —	+	India	Taygi 1982
		— » —	+	Thailand	Kiauta and Kiauta 1983
		— » —	+	India	Prasad and Thomas 1992
164.	<i>B. farinosa</i> Krueger, 1902	25(24A+X)	+	India	Dasgupta 1957
		— » —	+	India	Taygi 1982
		— » —	—	Thailand	Kiauta and Kiauta 1983
165.	<i>B. sobrina</i> (Rambur, 1842)	25(24A+X)	+	India	Ray Chaudhuri and Dasgupta 1949
		— » —	+	India	Taygi 1982
		— » —	+	Nepal	Kiauta and Kiauta 1982
166.	<i>Brachvmesia furcata</i> (Hagen, 1861)	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Argentina	Agopian and Mola 1988
		— » —	—	Brazil	Ferreira et al. 1979
		— » —	—	Brazil	Souza Bueno 1982
167.	<i>B. gravida</i> (Calvert, 1890)	25(24A+X)	+	USA	Cruden 1968 as <i>Cannacria gravida</i> (Calvert, 1890)
168.	<i>B. herbida</i> (Gundlach, 1889)	25(24A+X)	+	Jamaica	Cumming 1964 as <i>Cannacria herbida</i> (Gundlach, 1889)
169.	<i>Brachythemis contaminata</i> (Fabricius, 1793)	25(24A+X)	+	India	Asana and Makino 1935
		— » —	+	India	Makino 1935
		— » —	+	India	Kichijo 1942b
		— » —	+	India	Dasgupta 1957
		— » —	+	Nepal	Kiauta 1975
		— » —	+	India	Tyagi 1982
		— » —	+	Thailand	Kiauta and Kiauta 1983
170.	<i>B. lacustris</i> (Kirby, 1899)	25(24A+X)	+	Sudan	Wasscher 1985
171.	<i>Bradinopyga cornuta</i> Ris, 1911	25(24A+X)	+	Republic of South Africa	Boyes et al. 1980
172.	<i>B. geminata</i> (Rambur, 1842)	25(24A+X)	+	India	Dasgupta 1957
		— » —	+	India	Tyagi 1982
173.	<i>Brechmorhoga mendax</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
		— » —	—		
174.	<i>B. nubecula</i> (Rambur, 1842)	25(24A+X)	+	Bolivia	Cumming 1964
175.	<i>B. pertinax</i> (Hagen, 1861)	25(24A+X)	—	Bolivia	Cumming 1964 ( <i>B. p. peruviana</i> Ris, 1913)
176.	<i>Cannaphila vibex</i> (Hagen, 1861)	25(24A+X)	+	Bolivia	Cumming 1964
177.	<i>Celithemis amanda</i> (Hagen, 1861)	25(24A+X)	+	USA	Kiauta and Brink 1978
178.	<i>C. elisa</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
179.	<i>C. fasciata</i> Kirby, 1889	25(24A+X)	+	USA	Cruden 1968



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
180.	<i>C. ornata</i> (Rambur, 1842)	25(24A+X)	+	USA	Kiauta and Brink 1978
181.	<i>Crocothemis erythraea</i> (Brulle, 1832)	25(24A+X)	+	India	Dasgupta 1957
		— » —	+	Kenya	Kiauta 1969b
		— » —	+	Italy	Kiauta 1971a
		— » —	+	India	Prasad and Thomas 1992
		— » —	+	Republic of South Africa	Boyes et al. 1980
		— » —	+	India	Tyagi 1982
182.	<i>C. sanguinolenta</i> (Burmeister, 1839)	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
183.	<i>C. servilia</i> (Drury, 1773)	25(24A+X)	+	India	Asana and Makino 1935
		— » —	+	India	Makino 1935
		— » —	+	India	Kichijo 1942b
		— » —	+	India	Ray Chaudhuri and Dasgupta 1949
		— » —	+	Nepal	Kiauta 1975
		— » —	+	Philippines	Kiauta and Kiauta 1980b
		— » —	+	Nepal	Kiauta and Kiauta 1982
		— » —	+	India	Tyagi 1982
		— » —	+	Thailand	Kiauta and Kiauta 1983
		— » —	+	Japan	Katatani 1987
		— » —	+	Japan	Higashi and Kayano 1993
		— » —	+	Japan, Taiwan	Higashi et al. 2001
		24(22A+neo-XY)	+	Japan	Omura 1955 ( <i>C. s. mariannae</i> Kiauta, 1983)
		— » —	—	Japan	Kiauta 1983 ( <i>C. s. mariannae</i> )
		— » —	—	Japan	Katatani 1987 ( <i>C. s. mariannae</i> )
		— » —	—	Japan	Higashi et al. 2001 ( <i>C. s. mariannae</i> )
184.	<i>Dasythemis esmeralda</i> Ris, 1910	25(24A+X)	+	Bolivia	Cumming 1964
185.	<i>D. mincki</i> (Karsch, 1890)	25(24A+X)	+	Brazil	Souza Bueno 1982
186.	<i>D. venosa</i> (Burmeister, 1839)	25(24A+X)	+	Brazil	Kiauta and Boyes 1972
187.	<i>Diastatops intensa</i> Montgomery, 1940	25(24A+X)	+	Bolivia	Cumming 1964
188.	<i>D. obscura</i> (Fabricius, 1775)	25(24A+X)	+	Bolivia	Cumming 1964
189.	<i>D. pullata</i> (Burmeister, 1839)	23(22A+X)	+	Surinam	Kiauta 1979a
190.	<i>Diplacodes bipunctata</i> (Brauer, 1865)	25(24A+X)	+	Australia	Kiauta 1969b
		29(28A+X)	+		
191.	<i>D. haematodes</i> (Burmeister, 1839)	25(24A+X)	+	Australia	Kiauta 1969b
		23(22A+X)	—		
192.	<i>D. lefebvrei</i> (Rambur, 1842)	25(24A+X)	+	Madagascar	Kiauta 1968c, 1969b
193.	<i>D. nebulosa</i> (Fabricius, 1793)	25(24A+X)	+	India	Dasgupta 1957
		— » —	+	India	Kiauta and Kiauta 1982
		— » —	+	India	Tyagi 1982
194.	<i>D. trivialis</i> (Rambur, 1842)	25(24A+X)	+	India	Asana and Makino 1935
		— » —	+	India	Makino 1935
		— » —	+	India	Dasgupta 1957
		— » —	+	Australia	Kiauta 1969c
		— » —	+	Nepal	Kiauta 1975
		— » —	+	India	Tyagi 1982
195.	<i>Dythemis fugax</i> Hagen, 1861	25(24A+X)	+	USA	Cruden 1968
196.	<i>D. multipunctata</i> Kirby, 1894	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
197.	<i>D. rufinefris</i> (Burmeister, 1839)	25(24A+X)	+	Jamaica	Cumming 1964
198.	<i>D. velox</i> Hagen, 1861	25(24A+X)	+	Bolivia	Cumming 1964
		— » —	+	Peru	Kiauta and Boyes 1972
199.	<i>Elasmothermis cannaerioides</i> (Calvert, 1906)	21(20A+X)	—	Bolivia	Cumming 1964 as <i>Dythemis cannaerioides</i> Calvert, 1906
		23(22A+X)	+	Surinam	Kiauta 1979a as <i>D. cannaerioides</i>
		— » —	+	Brazil	Ferreira et al. 1979
200.	<i>E. williamsoni</i> (Ris, 1919)	22(20A+neo-XY)	—	Surinam	Kiauta 1979a as <i>Dythemis williamsoni</i> (Ris, 1919)
		25(24A+X)	—		



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
201.	<i>Erythemis attala</i> (Selys, 1857)	25(24A+X)	–	Bolivia	Cumming 1964
		– » –	+	Argentina	Agopian and Mola 1988
202.	<i>E. collocata</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
203.	<i>E. credula</i> (Hagen, 1861)	25(24A+X)	+	Surinam	Kiauta 1979a
204.	<i>E. haematogastra</i> (Burmeister, 1839)	25(24A+X)	–	Surinam	Kiauta 1979a
205.	<i>E. peruviana</i> (Rambur, 1842)	25(24A+X)	–	Surinam	Kiauta 1979a
206.	<i>E. plebeja</i> (Burmeister, 1839)	25(24A+X)	–	Bolivia	Cumming 1964
207.	<i>E. simplicicollis</i> (Say, 1839)	25(24A+X)	+	USA	Cruden 1968
208.	<i>E. vesiculosa</i> (Fabricius, 1775)	25(24A+X)	–	Bolivia	Cumming 1964 as <i>Lepthemis vesiculosa</i> (Fabricius, 1775)
		– » –	–	Surinam	Kiauta 1979a as <i>L. vesiculosa</i>
		– » –	+	Brasil	Ferreira et al. 1979 as <i>L. vesiculosa</i>
209.	<i>Erythrodiplax anomala</i> (Brauer, 1865)	25(24A+X)	+	Brazil	Souza Bueno 1982
210.	<i>E. atroterminala</i> Ris, 1911	25(24A+X)	+	Uruguay	Goni and Abenante 1982
		– » –	+	Argentina	Mola 1996
211.	<i>E. attenuata</i> (Kirby, 1889)	25(24A+X)	+	Surinam	Kiauta 1979a
		– » –	+	Brasil	Ferreira et al. 1979
212.	<i>E. basalis</i> (Kirby, 1897)	25(24A+X)	–	Bolivia	Cumming 1964
		– » –	+	Surinam	Kiauta 1979a ( <i>E. b. basalis</i> (Kirby, 1897))
		– » –	+	Brasil	Ferreira et al. 1979 ( <i>E. b. basalis</i> )
213.	<i>E. berenice</i> (Drury, 1770)	25(24A+X)	–	USA	Cruden 1968
		27(26A+X)	+	USA	Hung 1971
		25(24A+X)	+		
214.	<i>E. castanea</i> (Burmeister, 1839)	25(24A+X)	–	Bolivia	Cumming 1964
215.	<i>E. chromoptera</i> Borrer, 1942	23(22A+X)	+	Uruguay	Goni and Abenante 1982
216.	<i>E. cleopatra</i> Ris, 1911	25(24A+X)	+	Peru	Kiauta and Boyes 1972
217.	<i>E. connata</i> (Burmeister, 1839)	25(24A+X)	+	Chile	Kiauta and Boyes 1972 ( <i>E. c. connata</i> (Burmeister, 1839))
		– » –	+	USA	Kiauta and Brink 1978 ( <i>E. c. minuscula</i> (Rambur, 1842))
218.	<i>E. coralline</i> (Brauer, 1865)	25(24A+X)	+	Argentina	Mola 1996
219.	<i>E. famula</i> (Erichson, 1848)	25(24A+X)	+	Brazil	Souza Bueno 1982
220.	<i>E. fusca</i> (Rambur, 1842)	25(24A+X)	–	Bolivia	Cumming 1964 as <i>E. connata fusca</i> (Rambur, 1842)
		– » –	–	Guatemala	Cruden 1968 as <i>E. c. fusca</i>
		– » –	+	Surinam	Kiauta 1979a as <i>E. c. fusca</i>
		– » –	+	Brazil	Ferreira et al. 1979 as <i>E. c. fusca</i>
		– » –	+	Brazil	Souza Bueno 1982
		– » –	+	Argentina	Mola 1996
221.	<i>E. fervida</i> (Erichson, 1848)	25(24A+X)	+	Jamaica	Cumming 1964
222.	<i>E. justiniana</i> (Selys, 1857)	25(24A+X)	+	Jamaica	Cumming 1964
223.	<i>E. juliana</i> Ris, 1911	25(24A+X)	+	Brazil	Souza Bueno 1982
224.	<i>E. latimaculata</i> Ris, 1911	25(24A+X)	+	Surinam	Kiauta 1979a
		– » –	+	Brasil	Ferreira et al. 1979
225.	<i>E. hygaea</i> Ris, 1911	25(24A+X)	+	Argentina	Capitulo et al. 1991
		– » –	+	Argentina	Mola 1996
226.	<i>E. media</i> Borrer, 1942	21(20A+X)	+	Bolivia	Cumming 1964
		22(20A+XX)*	+	Brazil	Kiauta and Boyes 1972
		21(20A+X)	+	Surinam	Kiauta 1979a
		– » –	+	Brasil	Ferreira et al. 1979
		22(20A+neo-XY)	+	Argentina	Mola 1996
227.	<i>E. melanorubra</i> Borrer, 1942	25(24A+X)	+	Bolivia	Cumming 1964
		– » –	+	Venezuela	Kiauta and Boyes 1972
		– » –	+	Argentina	Capitulo et al. 1991
		– » –	+	Argentina	Mola 1996
228.	<i>E. minuscula</i> (Rambur, 1842)	25(24A+X)	+	USA	Kiauta and Brink 1978
		22(20A+neo-XY)	+	Argentina	Mola and Agopian 1985
229.	<i>E. nigricans</i> (Rambur, 1842)	25(24A+X)	+	Uruguay	Goni and Abenante 1982



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
229.	<i>E. nigricans</i> (Rambur, 1842)	– » –	+	Argentina	Mola 1996
		– » –	–	Argentina	De Gennaro 2004
		– » –	+	Argentina	De Gennaro et al. 2008
230.	<i>E. ochracea</i> (Burmeister, 1839)	25(24A+X)	+	Argentina	Mola 1996
231.	<i>E. paraguayensis</i> (Foerster, 1904)	23(22A+X)	+	Bolivia	Cumming 1964
		– » –	+	Surinam	Kiauta 1979a
232.	<i>E. umbrata</i> (Linnaeus, 1758)	25(24A+X)	+	Bolivia	Cumming 1964
		– » –	+	Dominica	Cruden 1968
		– » –	+	Surinam	Kiauta 1979a
		– » –	+	Brazil	Ferreira et al 1979
		– » –	+	Argentina	Mola 1996
233.	<i>E. unimaculata</i> (DeGeer, 1773)	25(24A+X)	+	Bolivia	Cumming 1964
		– » –	+	Surinam	Kiauta 1979a
234.	<i>Hydrobasileus croceus</i> (Brauer, 1867)	25(24A+X)	+	India	Prasad and Thomas 1992
235.	<i>Ladona julia</i> (Uhler, 1857)	25(24A+X)	+	USA	Cruden 1968
236.	<i>Lathrecista asiatica</i> (Fabricius, 1798)	25(24A+X)	+	India	Dasgupta 1957
		– » –	+	India	Tyagi 1982
237.	<i>Leucorrhinia albifrons</i> (Burmeister, 1839)	25(24A+X)	+	Former USSR	Makalowskaja 1940
238.	<i>L. dubia</i> (Van der Linden, 1825)	26(24A+XX)*	–	Finland	Oksala 1939a, 1945
		25(24A+X)	+	Russia	Kuznetsova et al. 2020b
239.	<i>L. frigida</i> Hagen, 1890	21(20A+X)	–	USA	Cruden 1968
		23(22A+X)	+		
240.	<i>L. glacialis</i> Hagen, 1890	25(24A+X)	+	USA	Cruden 1968
241.	<i>L. hudsonica</i> (Selys, 1850)	25(24A+X)	+	USA	Cruden 1968
		– » –	–		
242.	<i>L. intacta</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
		– » –	–		
243.	<i>L. pectoralis</i> (Charpentier, 1825)	26(24A+XX)*	–	Finland	Oksala 1945
244.	<i>L. proxima</i> Calvert, 1890	25(24A+X)	+	USA	Cruden 1968
245.	<i>L. rubicunda</i> (Linnaeus, 1857)	25(24A+X)	–	Finland	Oksala 1939a
		– » –	–	Former USSR	Makalowskaja 1940
		– » –	–	Russia	Kuznetsova et al. 2018
246.	<i>Libellula angelina</i> Selys, 1883	25(24A+X)	+	Japan	Oguma 1915, 1930
		– » –	+	Japan	Kichijo 1942a
247.	<i>L. auripennis</i> Burmeister, 1839	25(24A+X)	+	USA	Kiauta and Brink 1978
248.	<i>L. axilena</i> Westwood, 1837	23(22A+X)	–	USA	Cumming 1964
249.	<i>L. basalis</i> (Say, 1840)	25(24A+X)	–	USA	Smith 1916
250.	<i>L. composita</i> (Hagen, 1873)	25(24A+X)	+	USA	Cruden 1968
251.	<i>L. croceipennis</i> Selys, 1868	25(24A+X)	+	USA	Cruden 1968
252.	<i>L. cyanea</i> Fabricius, 1775	25(24A+X)	–	USA	Cruden 1968
253.	<i>L. depressa</i> Linnaeus, 1758	23(22A+X)	–	Belgium	Carnoy 1885
		– » –	–	England	Hogben 1921
		25(24A+X)	+	Austria	Kiauta 1968c, 1969b
		23(22A+X)	–		
		25(24A+X)	+	France	Kiauta 1973b
		– » –	+	Croatia	Francović and Jurečić 1986, 1989
		– » –	+	Russia	Perepelov et al. 1998
		– » –	+	Russia	Kuznetsova et al. 2018
254.	<i>L. flavida</i> Rambur, 1842	25(24A+X)	+	USA	Cruden 1968
255.	<i>L. forensis</i> Hagen, 1861	25(24A+X)	+	USA	Cruden 1968
256.	<i>L. fulva</i> Muller, 1764	25(24A+X)	+	Switzerland	Kiauta and Kiauta 1979
		27(26A+X)	+	Croatia	Francović and Jurečić 1986, 1989
257.	<i>L. insecta</i> Hagen, 1861	25(24A+X)	–	USA	Cumming 1964
		– » –	–	USA	Cruden 1968
258.	<i>L. luctuosa</i> Burmeister, 1839	25(24A+X)	–	USA	Smith 1916
259.	<i>L. pulchella</i> Drury, 1773	25(24A+X)	+	USA	Cruden 1968
		– » –	+	Canada	Kiauta 1969a
260.	<i>L. quadrimaculata</i> Linnaeus, 1758	25(24A+X)	+	Japan	Oguma 1915, 1930 ( <i>L. q. asahinai</i> Schmidt, 1957)



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
260.	<i>L. quadrimaculata</i> Linnaeus, 1758	25(24A+X)	+	Japan	Kichijo 1942d ( <i>L. q. asahinai</i> )
		— » —	+	Japan	Omura 1955 ( <i>L. q. asahinai</i> )
		— » —	+	Japan	Kiauta 1968b, c ( <i>L. q. asahinai</i> )
		— » —	+	Former USSR	Fuchsówna and Sawczyńska 1928 ( <i>L. q. quadrimaculata</i> Linnaeus, 1758)
		— » —	+	Finland	Oksala 1939a, b, 1945 ( <i>L. q. quadrimaculata</i> )
		— » —	+	Former USSR	Makalowskaja 1940 ( <i>L. q. quadrimaculata</i> )
		— » —	+	Netherlands	Kiauta 1968b, c ( <i>L. q. quadrimaculata</i> )
		— » —	+	USA	Cruden 1968 ( <i>L. q. quadrimaculata</i> )
		— » —	+	Russia	Perepelov et al. 1998 ( <i>L. q. quadrimaculata</i> )
		— » —	+	Russia	Kuznetsova et al. 2018 ( <i>L. q. quadrimaculata</i> )
261.	<i>L. saturata</i> Uhler, 1857	25(24A+X)	+	USA	Cruden 1968
262.	<i>L. semifasciata</i> Burmeister, 1839	25(24A+X)	+	USA	Cruden 1968
263.	<i>L. vibrans</i> Fabricius, 1793	25(24A+X)	+	USA	Cruden 1968
264.	<i>Lyriothemis pachygastra</i> (Selys, 1878)	25(24A+X)	—	Japan	Omura 1955
265.	<i>Macrothemis declivata</i> Calvert, 1909	23(22A+X)	+	Brazil	Kiauta and Boyes 1972
266.	<i>M. hemichlora</i> (Burmeister, 1839)	6(4A+neo-XY)	—	Bolivia	Cumming 1964
267.	<i>M. imitans</i> Karsch, 1890	25(24A+X)	+	Brazil	Kiauta and Boyes 1972 ( <i>M. i. imitans</i> Karsch, 1890)
268.	<i>M. mortoni</i> Ris, 1913	25(24A+X)	+	Bolivia	Cumming 1964
269.	<i>M. musiva</i> Calvert, 1898	25(24A+X)	+	Bolivia	Cumming 1964
270.	<i>Macrothemis</i> sp.	25(24A+X)	+	Argentina	Mola 2007
271.	<i>Miathyria artemis</i> (Selys, 1857)	25(24A+X)	+	Surinam	Kiauta 1979a
272.	<i>M. marcella</i> (Selys, 1857)	25(24A+X)	+	Bolivia	Cumming 1964
		— » —	+	Surinam	Kiauta 1979a
		— » —	+	Argentina	Mola and Agopian 1985
		— » —	+	Brazil	Ferreira et al. 1979
273.	<i>Micrathyria artemis</i> Ris, 1911	25(24A+X)	+	Brazil	Ferreira et al. 1979
		— » —	+	Brazil	Souza Bueno 1982
274.	<i>M. atra</i> (Martin, 1897)	25(24A+X)	+	Bolivia	Cumming 1964
275.	<i>M. catenata</i> Calvert, 1909	25(24A+X)	+	Brazil	Souza Bueno 1982
		— » —	+	Argentina	Mola 2007
276.	<i>M. didyma</i> (Selys, 1857)	25(24A+X)	+	Jamaica	Cumming 1964
277.	<i>M. exima</i> Kirby, 1897	25(24A+X)	+	Surinam	Kiauta 1979a
278.	<i>M. hagenii</i> Kirby, 1890	25(24A+X)	+	Jamaica	Cumming 1964
279.	<i>M. hesperis</i> Ris, 1911	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
		— » —	+	Argentina	Mola et al. 1999
280.	<i>M. hypodydima</i> Calvert 1906	23(22A+X)	+	Brazil	Souza Bueno 1982
		25(24A+X)	+	Argentina	Agopian and Mola 1988
281.	<i>M. iheringi</i> Santos, 1946	23(22A+X)	+	Bolivia	Cumming 1964
282.	<i>M. laevigata</i> Calvert, 1909	25(24A+X)	+	Bolivia	Cumming 1964
		— » —	+	Brazil	Kiauta and Boyes 1972
283.	<i>M. longifasciata</i> Calvert, 1909	24(22A+neo-XY)	—	Argentina	Agopian and Mola 1988
284.	<i>M. ocellata</i> (Martin, 1897)	25(24A+X)	+	Bolivia	Cumming 1964
					( <i>M. o. dentiens</i> Calvert, 1909)
285.	<i>M. spuria</i> (Selys, 1900)	25(24A+X)	+	Bolivia	Cumming 1964
		— » —	+	Argentina	Mola et al. 1999
286.	<i>M. stawiariskii</i> Santos, 1953	25(24A+X)	+	Brazil	Souza Bueno 1982
287.	<i>M. ungulata</i> Foerster, 1907	23(20A+X <sub>1</sub> X <sub>2</sub> Y)	—	Argentina	Mola et al. 1999
288.	<i>M. cf. eximia</i> Kirby, 1879	21(20A+X)	—	Bolivia	Cumming 1964
289.	<i>M. sp. (ungulata</i> Foerster, 1907-group)	23(22A+X)	—	Bolivia	Cumming 1964
290.	<i>Nannothemis bella</i> (Uhler, 1857)	25(24A+X)	+	USA	Cruden 1968
291.	<i>Nesciothemis farinosa</i> (Foerster, 1898)	25(24A+X)	+	Kenya	Kiauta 1969c
		— » —	+	Kenya	Wasscher 1985



Taxon	Karyotype formula 2n ♂	m-chromo somes	Country	References
292. <i>Nesogonia blackburni</i> (McLachlan, 1883)	25(24A+X)	+	Hawaii	Kiauta 1969d
293. <i>Neurothemis fulvia</i> (Drury, 1773)	25(24A+X)	+	Nepal	Kiauta 1974, 1975
294. <i>N. intermedia</i> (Rambur, 1842)	25(24A+X)	+	Nepal	Kiauta 1974, 1975 ( <i>N. i. intermedia</i> (Rambur, 1842))
	— » —	+	Nepal	Kiauta and Kiauta 1982 ( <i>N. i. degener</i> (Sel, 1842))
295. <i>N. terminata</i> Ris, 1911	25(24A+X)	+	Philippines	Kiauta and Kiauta 1980b
296. <i>N. tullia</i> (Drury, 1773)	28(26A+neo-XY)	+	India	Ray Chaudhuri and Dasgupta 1949
	— » —	+	India	Kiauta 1969a ( <i>N. t. tullia</i> (Drury, 1773))
	— » —	+	India	Tyagi 1982 ( <i>N. t. tullia</i> )
	25(24A+X)	+	Thailand	Kiauta and Kiauta 1983
297. <i>Oligoclada amphinome</i> Ris, 1919	25(24A+X)	+	Surinam	Kiauta 1979a
298. <i>O. laetitia</i> Ris, 1911	23(22A+X)	+	Argentina	Mola and Agopian 1985
	21(20A+X)	—	Brazil	Souza Bueno 1982
299. <i>O. monosticha</i> Borrer, 1931	23(22A+X)	+	Surinam	Kiauta 1979a
	— » —	+	Brazil	Ferreira et al. 1979
300. <i>O. pachystigma</i> Karsch, 1890	23(22A+X)	+	Brazil	Souza Bueno 1982
301. <i>Orthemis aequilibris</i> Calvert, 1909	12(10A+neo-XY)	—	Surinam	Kiauta 1979a
302. <i>O. ambinigra</i> Calvert, 1909	12(10A+neo-XY)	—	Argentina	Agopian and Mola 1984
303. <i>O. biolleyi</i> Calvert, 1906	23(22A+X)	+	Bolivia	Cumming 1964
304. <i>O. cultiformis</i> Calvert, 1906	23(22A+X)	+	Bolivia	Cumming 1964
	— » —	+	Surinam	Kiauta 1979a
	— » —	+	Brazil	Ferreira et al. 1979
	23(22A+X)	+	Argentina	Mola 2007
306. <i>O. ferruginea</i> (Fabricius, 1775)	10(8A+neo-XY)***	—	Bolivia	Cumming 1964
	23(22A+X)	—	USA	
	— » —	+	Guatemala, Dominica	Cruden 1968
	— » —	+	Peru	Kiauta 1969a, 1971c
	— » —	+	Peru	Kiauta and Boyes 1972
	23(22A+X)	+	Surinam	Kiauta 1979a
	25(24A+X)	+		
	23(22A+X)	+	Brazil	Ferreira et al. 1979
	23(22A+X)	—	Brazil, Argentina	Mola and Agopian 1985
	24(22A+XX)*	+		
307. <i>O. levis</i> Calvert, 1906	6(4A+neo-XY)***	—	Bolivia	Cumming 1964
	8(6A+neo-XY)***	—		
308. <i>O. nodiplaga</i> Karsch, 1891	41(40A+X)	—	Argentina	Agopian and Mola 1984
309. <i>Orthetrum abbotti</i> Calvert, 1892	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
310. <i>O. albistylum</i> (Selys, 1848)	25(24A+X)	+	Italy	Kiauta 1971a ( <i>O. a. albistylum</i> (Selys, 1848))
	— » —	+	Russia	Perepelov et al. 1998
	— » —	+	Japan	Oguma 1915, 1917, 1930 ( <i>O. a. speciosum</i> (Uhler, 1858))
	— » —	+	India	Kichijo 1942b ( <i>O. a. speciosum</i> )
	— » —	+	Japan	Omura 1955 ( <i>O. a. speciosum</i> )
311. <i>O. azureum</i> (Rambur, 1842)	25(24A+X)	+	Madagascar	Kiauta 1969b, c
312. <i>O. brachiale</i> (Beauvois, 1805)	21(20A+X)	—	Kenya	Kiauta 1969b, c
	25(24A+X)	+	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979 ( <i>O. b. brachiale</i> (Beauvois, 1805))
313. <i>O. brunneum</i> (Fonscolombe, 1837)	25(24A+X)	+	Italy	Kiauta 1971a
	— » —	+	Russia	Perepelov et al. 1998
314. <i>O. cancellatum</i> (Linnaeus, 1758)	25(24A+X)	+	Finland	Oksala 1939a
	— » —	+	India	Dasgupta 1957
	— » —	+	Netherlands	Kiauta 1969a, b
	— » —	+	India	Tyagi 1982
	— » —	+	Russia	Kuznetsova et al. 2018

Taxon	Karyotype formula 2n ♂	m-chromo somes	Country	References
315. <i>O. chrysostigma</i> (Burmeister, 1839)	25(24A+X)	+	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979
	— » —	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
	— » —	+	Kenya	Wasscher 1985
316. <i>O. coerulescens</i> (Fabricius, 1798)	25(24A+X)	+	Austria	Kiauta 1969c
	23(22A+X)	—		
	25(24A+X)	+	Italy	Kiauta 1971a
	27(26A+X)	+		
317. <i>O. glaucum</i> (Brauer, 1865)	25(24A+X)	+	India	Dasgupta 1957
	— » —	+	India	Tyagi 1978a, b
	— » —	+	India	Handa and Batra 1980
	— » —	+	India	Tyagi 1982
	— » —	+	India	Handa et al. 1984
	— » —	+	India	Walia and Sandhu 2002
	— » —	+	India	Kumari and Gautam 2017
318. <i>O. guineese</i> (Ris, 1909)	25(24A+X)	+	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979
319. <i>O. japonicum</i> (Uhler, 1858)	25(24A+X)	+	Japan	Oguma 1917, 1930 ( <i>O. j. internum</i> McLachlan, 1894)
	— » —	+	Japan	Kichijo 1942b ( <i>O. j. internum</i> )
	— » —	+	Japan	Omura 1955 ( <i>O. j. internum</i> )
	— » —	+	Nepal	Kiauta 1975 ( <i>O. j. internum</i> )
	— » —	+	Nepal	Kiauta and Kiauta 1976 ( <i>O. j. internum</i> )
320. <i>O. julia</i> Kirby, 1900	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980 ( <i>O. j. falsum</i> (Longfeild, 1955))
	— » —	+	Kenya	Wasscher 1985 ( <i>O. j. falsum</i> )
321. <i>O. luzonicum</i> (Brauer, 1868)	25(24A+X)	+	Nepal	Kiauta 1975
	— » —	+	Nepal	Kiauta and Kiauta 1982
	— » —	+	India	Thomas and Prasad 1981
	— » —	+	India	Prasad and Thomas 1992
322. <i>O. melania</i> (Selys, 1883)	25(24A+X)	+	Japan	Oguma 1917
	— » —	+	Japan	Omura 1955
	— » —	+	Russia	Perepelov 2003
323. <i>O. monardi</i> (Schmidt, 1951)	25(24A+X)	+	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979
324. <i>O. poecilops</i> (Ris, 1916)	25(24A+X)	+	Japan	Suzuki et al. 1991 ( <i>O. p. miyajimaensis</i> Yuki et Doi, 1938)
325. <i>O. pruinosum</i> (Burmeister, 1839)	25(24A+X)	+	India	Dasgupta 1957 ( <i>O. p. neglectum</i> (Rambur, 1842))
	— » —	+	Taiwan	Kiauta 1969a, c ( <i>O. p. neglectum</i> )
	— » —	+	India	Tyagi 1982 ( <i>O. p. neglectum</i> )
	— » —	+	India	Prasad and Thomas 1992 ( <i>O. p. neglectum</i> )
	— » —	+	India	Tyagi 1978a, b ( <i>O. p. neglectum</i> )
	— » —	+	Nepal	Kiauta and Kiauta 1982 ( <i>O. p. neglectum</i> )
	— » —	+	India	Walia and Sandhu 2002 ( <i>O. p. neglectum</i> )
	— » —	+	India	Kumari and Gautam 2017 ( <i>O. p. neglectum</i> )
326. <i>O. sabina</i> (Drury, 1773)	25(24A+X)	+	India	Asana and Makino 1935
	— » —	+	India	Makino 1935
	— » —	+	India	Kichijo 1942b
	— » —	+	India	Ray Chaudhuri and Dasgupta 1949
	— » —	+	Nepal	Kiauta 1975



Taxon	Karyotype formula 2n ♂	m-chromo somes	Country	References
326. <i>O. sabina</i> (Drury, 1773)	— » —	+	India	Tyagi 1982
	— » —	+	India	Prasad and Thomas 1992
	— » —	+	India	Walia and Sandhu 2002 ( <i>O. s. sabina</i> (Drury, 1773))
327. <i>O. taeniolatum</i> (Schneider, 1845)	25(24A+X)	+	Greece	Kiauta 1972a
	— » —	+	Nepal	Kiauta 1975
	— » —	+	India	Tyagi 1978a, b
	— » —	+	India	Handa and Batra 1980
	— » —	+	India	Tyagi 1982
	— » —	+	India	Handa et al. 1984
	— » —	+	India	Thomas and Prasad 1986
	— » —	+	India	Walia and Sandhu 2002a
	— » —	+	India	Walia et al. 2015
328. <i>O. testaceum</i> (Burmeister, 1839)	25(24A+X)	+	Nepal	Kiauta and Kiauta 1982
329. <i>O. triangulare</i> (Selys, 1878)	25(24A+X)	+	Japan	Omura 1955 ( <i>O. t. melania</i> (Selys, 1883))
	— » —	+	Taiwan	Kiauta 1969a, b ( <i>O. t. triangulare</i> (Selys, 1878))
	— » —	+	Nepal	Kiauta 1975 ( <i>O. t. triangulare</i> )
	— » —	+	India	Tyagi 1978a, b ( <i>O. t. triangulare</i> )
	— » —	+	India	Handa and Batra 1980 ( <i>O. t. triangulare</i> )
	— » —	+	India	Tyagi 1982 ( <i>O. t. triangulare</i> )
	— » —	+	India	Walia and Sandhu 2002 ( <i>O. t. triangulare</i> )
330. <i>Pachydiplax longipennis</i> (Burmeister, 1839)	25(24A+X)	—	USA	Cumming 1964
	— » —	+	USA	Cruden 1968
	— » —	+	USA	Kiauta and Brink 1978
331. <i>Palpopleura jucunda</i> Rambur, 1842	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
332. <i>P. lucia</i> (Drury, 1773)	25(24A+X)	+	Burkina Faso (Former Voltaic Republic)	Kiauta and Ochssée 1979 ( <i>P. l. portia</i> (Drury, 1773))
	— » —	+	Kenya	Wasscher 1985 ( <i>P. l. portia</i> )
333. <i>P. sexmaculata</i> (Fabricius, 1787)	25(24A+X)	+	Nepal	Kiauta 1974, 1975
	— » —	+	India	Tyagi 1982 ( <i>P. s. sexmaculata</i> (Fabricius, 1787))
334. <i>Pantala flavescens</i> (Fabricius, 1798)	25(24A+X)	+	India	Asana and Makino 1935
	— » —	+	India	Makino 1935
	— » —	+	India	Kichijo 1942b
	— » —	+	India	Dasgupta 1957
	— » —	+	India	Seshachar and Bagga 1963
	— » —	+	Bolivia	Cumming 1964
	— » —	+	Madagascar	Kiauta 1969b
	— » —	+	Surinam	Kiauta 1979a
	— » —	+	Brazil	Ferreira et al. 1979
	— » —	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
	— » —	+	Brazil	Souza Bueno 1982
	— » —	+	Argentina	Agopian and Mola 1988
	— » —	+	India	Prasad and Thomas 1992
	— » —	+	Russia	Perepelov and Bugrov 2001b
	23(22A+X)	+	India	Walia et al. 2011
335. <i>P. hymenaea</i> (Say, 1836)	25(24A+X)	+	Bolivia	Cumming 1964
	— » —	+	USA	Cruden 1968
336. <i>Perithemis cornelia</i> Ris, 1910	25(24A+X)	—	Bolivia	Cumming 1964
337. <i>P. domitia</i> (Drury, 1773)	25(24A+X)	+	Jamaica	Cumming 1964
338. <i>P. electra</i> Ris, 1928	25(24A+X)	—	Bolivia	Cumming 1964
339. <i>P. icteroptera</i> (Selys in Sagra, 1857)	25(24A+X)	+	Argentina	Mola and Agopian 1985

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
340.	<i>P. lais</i> (Petry, 1834)	17(16A+X)	—	Bolivia	Cumming 1964
		— » —	—	Surinam	Kiauta 1979a
		— » —	—	Brazil	Ferreira et al. 1979
341.	<i>P. mooma</i> Kirby, 1889	25(24A+X)	+	Bolivia	Cumming 1964
		— » —	+	Surinam	Kiauta 1979a
		— » —	—	Brazil	Ferreira et al. 1979
		— » —	+	Argentina	Mola and Agopian 1985
342.	<i>P. tenera</i> (Say, 1839)	25(24A+X)	+	USA	Kiauta and Brink 1978
343.	<i>P. seminole</i> Calvert, 1907	25(24A+X)	+	USA	Cumming 1964
344.	<i>Perithemis</i> sp.	25(24A+X)	—	Bolivia	Cumming 1964
345.	<i>Planiplax erythropyga</i> (Karsch, 1891)	25(24A+X)	+	Argentina	Mola et al. 1999
		— » —	+	— » —	De Gennaro 2004
346.	<i>P. sanguiniventris</i> (Calvert, 1907)	25(24A+X)	+	USA	Cruden 1968
347.	<i>Plathemis lydia</i> (Drury, 1773)	25(24A+X)	+	USA	McGill 1907
		— » —	+	USA	Cruden 1968
348.	<i>Potamarcha congener</i> (Rambur, 1842)	25(24A+X)	+	India	Asana and Makino 1935 as <i>P. obscura</i> (Rambur, 1842)
		— » —	+	India	Makino 1935 as <i>P. obscura</i>
		— » —	+	India	Kichijo 1942b as <i>P. obscura</i>
		— » —	+	India	Dasgupta 1957 as <i>P. obscura</i>
		— » —	+	India	Tyagi 1982 as <i>P. obscura</i>
		— » —	+	India	Prasad and Thomas 1992
349.	<i>Pseudothemis zonata</i> (Burmeister, 1839)	24(22A+neo-XY)	—	Japan	Omura 1955
350.	<i>Pseudotranea prateri</i> Fraser, 1920	25(24A+X)	+	Nepal	Kiauta 1974, 1975
351.	<i>Rhodopygia cardinalis</i> (Erichson, 1848)	25(24A+X)	+	Bolivia	Cumming 1964
352.	<i>R. geijskesi</i> Belle, 1964	25(24A+X)	+	Surinam	Kiauta 1979a
353.	<i>Rhodothemis rufa</i> (Rambur, 1842)	25(24A+X)	+	India	Prasad and Thomas 1992
354.	<i>Rhyothemis fuliginosa</i> Selys, 1883	25(24A+X)	+	Japan	Toyoshima and Hirai 1953
		— » —	+	Japan	Omura 1955
		— » —	+	Japan	Hirai 1956
		25(24A+X)	+	Japan	Kiauta 1969c
		23(22A+X)	+		
355.	<i>R. variegata</i> (Linnaeus et Johansson, 1763)	25(24A+X)	+	India	Ray Chaudhuri and Dasgupta 1949
		— » —	+	Nepal	Kiauta 1975
356.	<i>Scapanea frontalis</i> (Burmeister, 1839)	25(24A+X)	+	Jamaica	Cumming 1964
357.	<i>Sympetrum commixtum</i> (Selys, 1884)	25(24A+X)	—	India	Tyagi 1978a, b, 1982
358.	<i>S. corruptum</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968 as <i>Tarnetrum corruptum</i> (Hagen, 1861)
		— » —	+	USA	Kiauta 1969a, c as <i>T. corruptum</i>
359.	<i>S. costiferum</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
360.	<i>S. croceolum</i> (Selys, 1840)	25(24A+X)	+	Russia	Perepelov 2003
361.	<i>S. danae</i> (Sulzer, 1776)	25(24A+X)	+	Former USSR	Makalowskaja 1940
		— » —	+	Finland	Oksala 1945
		— » —	+	USA	Cruden 1968
		— » —	+	Russia	Perepelov 2003
		— » —	+	Russia	Kuznetsova et al. 2018
362.	<i>S. eroticum</i> (Selys, 1883)	21(20A+X)	—	Japan	Kichijo 1942b, c
		— » —	—	Japan	Hirai 1956
		— » —	—	Japan	Kiauta 1969c
363.	<i>S. flaveolum</i> (Linnaeus, 1758)	25(24A+X)	+	Former USSR	Makalowskaja 1940
		— » —	+	Russia	Perepelov 2003
364.	<i>S. fonscolombii</i> (Selys, 1840)	25(24A+X)	+	Russia	Perepelov 2003
365.	<i>S. frequens</i> (Selys, 1883)	23(22A+X)	—	Japan	Oguma 1917, 1930
		— » —	—	Japan	Kichijo 1942a, b
		— » —	—	Japan	Kiauta 1969c
366.	<i>S. infuscatum</i> (Selys, 1883)	25(24A+X)	+	Russia	Perepelov 2003
367.	<i>S. internum</i> Montgomery, 1943	27(26A+X)	+	Canada	Kiauta 1973a
368.	<i>S. madidum</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
368.	<i>S. madidum</i> (Hagen, 1861)	— » —	+	Canada	Kiauta 1973a
369.	<i>S. meridionale</i> (Selys, 1841)	25(24A+X)	+	Switzerland	Kiauta 1966
370.	<i>S. obtrusum</i> (Hagen, 1867)	25(24A+X)	+	USA	Cruden 1968
371.	<i>S. parvulum</i> Bartenev, 1912	25(24A+X)	+	Japan	Kiauta 1968c
372.	<i>S. pedemontanum</i> Müller in Allioni, 1766	25(24A+X)	+	Japan	Oguma 1917, 1930 ( <i>S. p. elatum</i> (Selys, 1872))
		— » —	+	Japan	Kichijo 1942b ( <i>S. p. elatum</i> )
		— » —	+	Japan	Kiauta and Brink 1975 ( <i>S. p. elatum</i> )
		— » —	+	Switzerland	Kiauta and Brink 1975 ( <i>S. p. pedemontanum</i> (Müller, 1766))
		— » —	+	Russia	Perepelov et al. 1998 ( <i>S. p. pedemontanum</i> )
		— » —	+	Russia	Perepelov and Bugrov 2001b
373.	<i>S. rubicundulum</i> (Say, 1839)	25(24A+X)	+	USA	Cruden 1968
374.	<i>S. sanguineum</i> (Müller, 1764)	25(24A+X)	+	Italy	Kiauta 1971a
		— » —	+	Russia	Perepelov and Bugrov 2001b
375.	<i>S. semicinctum</i> (Say, 1839)	25(24A+X)	+	USA	Smith 1916
		— » —	+	USA	Cruden 1968
376.	<i>S. striolatum</i> (Charpentier, 1840)	25(24A+X)	—	Luxembourg	Kiauta 1966
377.	<i>S. vicinum</i> (Hagen, 1861)	25(24A+X)	+	USA	Cruden 1968
378.	<i>S. vulgatum</i> (Linnaeus, 1758)	25(24A+X)	+	Netherland	Kiauta 1972c
		— » —	+	Russia	Perepelov 2003
		— » —	+	Russia	Kuznetsova et al. 2018
379.	<i>Tarnetrum</i> <i>illotum</i> (Hagen, 1861)	25(24A+X)	+	Jamaica	Cumming 1964
		— » —	+	USA	Cruden 1968
380.	<i>Tauriphila</i> <i>australis</i> (Hagen, 1867)	25(24A+X)	+	Bolivia	Cumming 1964
381.	<i>T. azteca</i> Calvert, 1906	25(24A+X)	+	Mexico	Cruden 1968
382.	<i>T. risi</i> Martin 1896	25(24A+X)	+	Argentina, Uruguay	Mola and Agopian 1985
383.	<i>Tholymis</i> <i>citrina</i> Hagen, 1867	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
384.	<i>Th. tillagra</i> (Fabricius, 1798)	25(24A+X)	+	India	Prasad and Thomas 1992
		— » —	+	Nepal	Kiauta and Kiauta 1982
		— » —	+	Thailand	Kiauta and Kiauta 1983
385.	<i>Tramea</i> <i>abdominalis</i> (Rambur, 1842)	25(24A+X)	—	Bolivia	Cumming 1964
386.	<i>T. basilaris</i> (Palisot de Beauvois, 1817)	25(24A+X)	+	India	Das 1956 ( <i>T. b. burmeisteri</i> (Kirby, 1889))
		— » —	+	India	Dasgupta 1957 ( <i>T. b. burmeisteri</i> )
		— » —	+	Nepal	Kiauta and Kiauta 1982 ( <i>T. b. burmeisteri</i> )
		— » —	+	India	Prasad and Thomas 1992 ( <i>T. b. burmeisteri</i> )
387.	<i>T. binotata</i> (Rambur, 1842)	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	—	Brazil	Ferreira et al. 1979
388.	<i>T. carolina</i> (Linnaeus, 1763)	25(24A+X)	—	USA	Cumming 1964
		— » —	—	USA	Cruden 1968
389.	<i>T. cophysa</i> (Hagen, 1867)	25(24A+X)	+	Bolivia	Cumming 1964
390.	<i>T. lacerata</i> (Hagen, 1861)	25(24A+X)	—	USA	Cruden 1968
391.	<i>T. limbata</i> (Desjardins, 1832)	25(24A+X)	+	India	Asana and Makino 1935
		— » —	+	India	Makino 1935
		— » —	+	India	Kichijo 1942b
392.	<i>T. virginia</i> (Rambur, 1842)	25(24A+X)	+	India	Oguma and Asana 1932
		— » —	+	India	Kichijo 1942b
		— » —	+	India	Dasgupta 1957
393.	<i>Trithemis</i> <i>annulata</i> (Palisot de Beauvois, 1805)	25(24A+X)	—	Republic of South Africa	Boyes et al. 1980
		— » —	+	Kenya	Wasscher 1985
394.	<i>T. arteriosa</i> (Burmeister, 1839)	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
395.	<i>T. atra</i> Pinhey, 1961	25(24A+X)	+	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979
396.	<i>T. aurora</i> (Burmeister, 1839)	25(24A+X)	+	India	Oguma and Asana 1932
		— » —	+	Nepal	Kiauta 1975
		— » —	+	India	Tyagi 1982
397.	<i>T. dorsalis</i> (Rambur, 1842)	25(24A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
398.	<i>T. festiva</i> (Rambur, 1842)	25(24A+X)	+	Nepal	Kiauta 1974, 1975
		— » —	+	India	Tyagi 1982
		— » —	+	India	Prasad and Thomas 1992
399.	<i>T. furva</i> Karsch, 1899	25(24A+X)	+	Sudan	Wasscher 1985
400.	<i>T. imiata</i> Pinhey, 1961	25(24A+X)	—	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979
401.	<i>T. kirbyi</i> Selys, 1891	25(24A+X)	—	Burkina Faso (Former Voltiac Republic)	Kiauta and Ochssée 1979 ( <i>T. k. ardens</i> Gerstaecker, 1891)
		— » —	+	Kenya	Wasscher 1985 ( <i>T. k. ardens</i> )
402.	<i>T. pallidinervis</i> (Kirby, 1889)	25(24A+X)	+	India	Asana and Makino 1935
		— » —	+	India	Makino 1935
		— » —	+	India	Kichijo 1942b
		— » —	+	India	Dasgupta 1957
		— » —	+	Philippines	Kiauta and Kiauta 1980b
403.	<i>T. werneri</i> Ris, 1912	25(24A+X)	+	Kenya	Wasscher 1985
404.	<i>Uracis imbuta</i> (Burmeister, 1839)	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
405.	<i>U. oviposatrix</i> Calvert, 1909	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	—	Brazil	Ferreira et al. 1979
406.	<i>Urothemis edwardsi</i> (Selys, 1849)	25(24A+X)	+	Sudan	Wasscher 1985
407.	<i>U. signata</i> (Rambur, 1842)	25(24A+X)	+	India	Das 1956 ( <i>U. s. signata</i> (Rambur, 1842))
		— » —	+	India	Dasgupta 1957 ( <i>U. s. signata</i> )
		— » —	+	Nepal	Kiauta 1975
		— » —	+	India	Prasad and Thomas 1992
408.	<i>Zenithoptera fasciata</i> (Linnaeus, 1758)	25(24A+X)	+	Surinam	Kiauta 1979a
409.	<i>Z. lanei</i> Santos, 1941	25(24A+X)	+	Surinam	Kiauta 1979a
		— » —	+	Brazil	Ferreira et al. 1979
410.	<i>Z. viola</i> Ris, 1910	25(24A+X)	+	Bolivia	Cumming 1964
411.	<i>Zygonyx iris</i> Kirby, 1900	23(22A+X)	+	Thailand	Kiauta and Kiauta 1983 ( <i>Z. i. malayanus</i> (Laidlaw, 1902))
412.	<i>Z. torrida</i> (Kirby, 1889)	25(24A+X)	+	India	Tyagi 1978a, b
413.	<i>Zyxomma petiolatum</i> (Rambur, 1842)	25(24A+X)	+	India	Prasad and Thomas 1992
CORDULEGASTROIDEA					
Chlorogomphidae					
414.	<i>Watanabeopetalia atkinsoni</i> (Selys, 1878)	25(24A+X)	+	India	Walia and Chahal 2019
Cordulegastridae					
415.	<i>Anotogaster basalis</i> Selys, 1854	23(22A+X)	—	India	Sandhu and Malhotra 1994b
416.	<i>A. kuchenbeiseri</i> (Förster, 1899)	25(24A+X)	+	China	Zhu and Wu 1986
417.	<i>A. sieboldii</i> (Selys, 1854)	25(24A+X)	+	Japan	Oguma 1930
		— » —	+	Japan	Kichijo 1942a
		— » —	+	Japan	Kiauta 1969a
		— » —	+	Russia	Perepelov et al. 2001
418.	<i>Cordulegaster boltoni</i> (Donovan, 1807)	25(24A+X)	+	Finland	Oksala 1939a, b
		— » —	—	Austria	Kichijo 1942a
		— » —	+	Sweden	Kiauta 1968d, e, 1969a
419.	<i>C. brevistigma</i> Selys, 1854	25(24A+X)	+	India	Walia and Chahal 2019
420.	<i>C. diastatops</i> (Selys, 1854)	25(24A+X)	+	USA	Cruden 1968
421.	<i>C. dorsalis</i> Hagen, 1857	25(24A+X)	+	USA	Cruden 1968



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
422.	<i>C. maculata</i> Selys, 1854	25(24A+X)	+	USA	Cruden 1968
423.	<i>Neallogaster hermionae</i> (Fraser, 1927)	25(24A+X)	+	Nepal	Kiauta and Kiauta 1976
ZYGOPTERA					
LESTOIDEA					
Lestidae					
424.	<i>Austrolestes colensonis</i> (White, 1846)	25(24A+X)	+	New Zealand	Jensen 1980
425.	<i>Chalcolestes viridis</i> (Van der Linden, 1825)	25(24A+X)	+	Netherlands	Kiauta 1969a
426.	<i>Indolestes cyaneus</i> (Selys, 1862)	25(24A+X)	+	Nepal	Kiauta and Kiauta 1976 as <i>I. cyanea</i> (Selys, 1862)
427.	<i>Lestes barbarus</i> (Fabricius, 1798)	25(24A+X)	+	Former Yugoslavia	Kiauta 1972a
428.	<i>L. congener</i> Hagen, 1861	25(24A+X)	+	USA	Cruden 1968
429.	<i>L. disjunctus</i> Selys, 1862	25(24A+X)	–	USA	Cruden 1968
430.	<i>L. dorothea</i> Fraser, 1924	25(24A+X)	+	Nepal	Kiauta 1974, 1975
431.	<i>L. dryas</i> Kirby, 1890	25(24A+X)	–	USA	Cruden 1968
		– » –	+	Russia	Perepelov and Bugrov 2001b
432.	<i>L. forcipatus</i> Rambur, 1842	21(20A+X)	–	USA	Cruden 1968
433.	<i>L. forcicula</i> Rambur, 1842	25(24A+X)	+	Jamaica	Cumming 1964
434.	<i>L. paulistus</i> Calvert, 1909	25(24A+X)	+	Brazil	Souza Bueno 1982
435.	<i>L. rectangularis</i> Say, 1839	25(24A+X)	+	USA	Cruden 1968
436.	<i>L. similatrix</i> McLachlan, 1895	25(24A+X)	+	Madagascar	Kiauta 1969b
437.	<i>L. sponsa</i> (Hansemann, 1823)	25(24A+X)	–	Former USSR	Makalowskaja 1940
		– » –	+	Japan	Kichijo 1941, 1942a, d, e
		– » –	+	Russia	Perepelov and Bugrov 2001b
438.	<i>L. stultus</i> Hagen, 1861	25(24A+X)	+	USA	Cruden 1968
439.	<i>L. vidua</i> Hagen, 1861	25(24A+X)	+	USA	Cumming 1964
440.	<i>L. vigilax</i> Selys, 1862	19(18A+X)	–	USA	Kiauta and Brink 1978
441.	<i>L. virens</i> Charpentier, 1825	25(24A+X)	+	Netherlands	Kiauta 1969a ( <i>L. v. vestalis</i> Rambur, 1842)
442.	<i>Sympecma fusca</i> (Van der Linden, 1823)	25(24A+X)	+	Japan	Kichijo 1941, 1942d, e
443.	<i>S. paedisca</i> (Brauer, 1877)	25(24A+X)	+	Netherlands	Kiauta and Kiauta-Brink 1975 ( <i>S. annulata braueri</i> (Bianchi, 1904))
		– » –	+	Russia	Perepelov 2003 ( <i>S. a. braueri</i> )
Synlestidae					
444.	<i>Megalestes major</i> Selys, 1862	25(24A+X)	–	Nepal	Kiauta 1974, 1975
PLATYSTICTOIDEA					
Platystictidae					
445.	<i>Drepanosticta</i> sp.	25(24A+X)	–	Nepal	Kiauta and Kiauta 1976
446.	<i>Drepanosticta</i> sp.	25(24A+X)	–	India	Tyagi 1978a, b
447.	<i>Palaemnema paulina</i> (Drury, 1773)	25(24A+X)	+	Costa Rica	Cumming 1964
448.	<i>Protosticta</i> sp.	25(24A+X)	–	Tailand	Kiauta and Kiauta 1983
CALOPTERYGOIDEA					
Calopterygidae					
449.	<i>Atrocalopteryx atrata</i> (Selys, 1853)	25(24A+X)	+	Japan	Oguma 1930 as <i>Calopteryx atrata</i> Selys, 1853
		– » –	+	Japan	Kichijo 1942d as <i>C. atrata</i>
		– » –	+	Japan	Omura 1957 as <i>C. atrata</i>
450.	<i>Calopteryx aequabilis</i> Say, 1839	25(24A+X)	+	USA	Cruden 1968
451.	<i>C. cornelia</i> (Selys, 1853)	25(24A+X)	+	Japan	Oguma 1930 as <i>Anaciagrion cornelia</i> (Selys, 1853)
		– » –	+	Japan	Kichijo 1942a as <i>A. cornelia</i>
452.	<i>C. dimidiata</i> Burmeister, 1839	25(24A+X)	+	USA	Kiauta and Brink 1978
453.	<i>C. japonica</i> Selys, 1869	25(24A+X)	+	Japan	Kichijo 1942a
		– » –	+	Japan	Hirai 1956
		– » –	+	Japan	Omura 1957
		– » –	+	Japan	Kiauta 1968e, f
454.	<i>C. maculata</i> (Beauvois, 1805)	25(24A+X)	+	USA	Cumming 1964a
		– » –	+	USA	Cruden 1968
455.	<i>C. splendens</i> (Harris, 1780)	25(24A+X)	+	Turkey	Kiauta 1972a ( <i>C. s. amasina</i> Bartenev, 1912)

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
455.	<i>C. splendens</i> (Harris, 1780)	— » —	+	Italy	Kiauta 1971a ( <i>C. s. caprai</i> Conci, 1956)
		— » —	—	Former USSR	Makalowskaja 1940 ( <i>C. s. splendens</i> (Harris, 1782))
		— » —	—	Finland	Oksala 1945 ( <i>C. s. splendens</i> )
		— » —	—	Germany	Kiauta 1969a, 1971b ( <i>C. s. splendens</i> )
		— » —	—	France	Kiauta 1973b ( <i>C. s. splendens</i> )
		— » —	—	Russia	Perepelov et al. 1998 ( <i>C. s. splendens</i> )
		— » —	+	Russia	Kuznetsova et al. 2020b
456.	<i>C. virgo</i> (Linnaeus, 1758)	25(24A+X)	+	Spain	Kiauta 1971b ( <i>C. v. meridionalis</i> Selys, 1873)
		27(26A+X)	+		
		25(24A+X)	+	Slovenija	Kiauta 1967a, 1968b, c ( <i>C. v. padana</i> Conci, 1956)
		— » —	+	Austria	Kiauta 1967a, 1968b, c ( <i>C. v. padana</i> )
		— » —	—	Belgium	Carnoy 1885 ( <i>C. v. virgo</i> (Linnaeus, 1758))
		— » —	+	Finland	Oksala 1939 ( <i>C. v. virgo</i> )
		— » —	+	Former USSR	Makalowskaja 1940 ( <i>C. v. virgo</i> )
		— » —	+	Germany, Luxembourg	Kiauta 1968e, f ( <i>C. v. virgo</i> )
		— » —	+	Netherlands	Kiauta 1972c ( <i>C. v. virgo</i> )
		— » —	+	Russia	Kuznetsova et al. 2020b
457.	<i>Hetaerina americana</i> (Fabricius, 1798)	25(24A+X)	+	USA	Cumming 1964
		— » —	—	USA	Cruden 1968
458.	<i>H. charca</i> Calvert, 1909	25(24A+X)	+	Bolivia	Cumming 1964
459.	<i>H. longipes</i> (Hagen in Selys, 1853)	25(24A+X)	+	Brazil	Souza Bueno 1982 as <i>H. carnifex</i> Hagen in Selys, 1853
		— » —	+	Brazil	Agopian and Mola 1984 as <i>H. carnifex</i>
460.	<i>H. rosea</i> Selys, 1853	27(26A+X)	+	Bolivia	Cumming 1964
		— » —	+	Bolivia	Kiauta 1969c
		25(24A+X)	—	Brazil	Ferreira et al. 1979
		27(26A+X)	+		
461.	<i>H. sanguinea</i> Selys, 1853	25(24A+X)	—	Bolivia	Cumming 1964
462.	<i>H. titia</i> (Drury, 1773)	25(24A+X)	+	USA	Cumming 1964
		— » —	+	Mexico	Kiauta 1970a as <i>H. tricolor</i> (Burmeister, 1839)
463.	<i>H. vulnerata</i> (Selys, 1853)	25(24A+X)	+	Mexico	Kiauta 1970a
464.	<i>Matrona basilaris</i> Selys, 1853	25(24A+X)	—	Taiwan	Kiauta 1968c
465.	<i>Mnais costalis</i> Selys, 1869	25(24A+X)	+	Japan	Oguma 1930
		— » —	+	Japan	Kichijo 1942a
466.	<i>M. pruinosa</i> Selys, 1853	25(24A+X)	+	Japan	Oguma 1930 as <i>M. strigata</i> Selys, 1853
		— » —	+	Japan	Kichijo 1942a as <i>M. strigata</i>
		— » —	+	Japan	Omura 1957 as <i>M. strigata</i>
467.	<i>Neurobasis chinensis</i> (Linnaeus, 1758)	23(22A+X)	—	Nepal	Kiauta 1975 ( <i>N. c. chinensis</i> (Linnaeus, 1758))
		25(24A+X)	—		
		23(22A+X)	—	India	Tyagi 1978b ( <i>N. c. chinensis</i> )
		— » —	+	Nepal	Kiauta and Kiauta 1982 ( <i>N. c. chinensis</i> )
		— » —	—	Thailand	Kiauta and Kiauta 1983 ( <i>N. c. chinensis</i> )
		— » —	+	India	Walia and Sandhu 2002 ( <i>N. c. chinensis</i> )
		— » —	—	India	Walia et al. 2016 ( <i>N. c. chinensis</i> )
		— » —	—	India	Walia and Katnoria 2018 ( <i>N. c. chinensis</i> )
468.	<i>Phaon iridipennis</i> (Burmeister, 1839)	25(24A+X)	+	Republic of South Africa	Boyes et al. 1980
<b>Chlorocyphidae</b>					
469.	<i>Aristocypha fenestrella</i> Rambur, 1842	23(22A+X)	—	Thailand	Kiauta and Kiauta 1983 as <i>Rhinocypha fenestrella</i> Rambur, 1842



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
470.	<i>A. quadrimaculata</i> (Selys, 1853)	23(22A+X)	+	India	Chatterjee and Kiauta 1973 as <i>Rhinocypha quadrimaculata</i> Selys, 1853
		— » —	+	Nepal	Kiauta and Kiauta 1982 as <i>Rh. quadrimaculata</i>
471.	<i>A. trifasciata</i> (Selys, 1853)	23(22A+X)	—	India	Tyagi 1978a, b as <i>Rhinocypha trifasciata</i> Selys, 1853
		— » —	+	Nepal	Kiauta and Kiauta 1982 as <i>Rh. trifasciata</i>
472.	<i>Helioocypha biforata</i> (Selys, 1859)	23(22A+X)	—	India	Tyagi 1978 a, b as <i>Rhinocypha biforata beesoni</i> Selys, 1859
473.	<i>H. biseriata</i> (Selys, 1859)	23(22A+X)	—	Thailand	Kiauta and Kiauta 1983 as <i>Rhinocypha b. biforata</i> Selys, 1859
474.	<i>Libellago lineata</i> (Burmeister, 1839)	23(22A+X)	—	India	Walia et al. 2018 ( <i>L. l. lineata</i> (Burmeister, 1839))
		25(24A+X)	—		
475.	<i>Paracypha unimaculata</i> (Selys, 1879)	23(22A+X)	+	Nepal	Kiauta 1974, 1975 as <i>Rhinocypha unimaculata</i> Selys, 1879
		— » —	+	Nepal	Kiauta and Kiauta 1982 as <i>Rh. unimaculata</i>
476.	<i>Rhinocypha colorata</i> Selys, 1869	23(22A+X)	—	Philippines	Kiauta and Kiauta 1980b
		25(24A+X)	—		
477.	<i>Vestalis gracilis</i> (Rambur, 1842)	25(24A+X)	+	Thailand	Kiauta and Kiauta 1983
Polythoridae					
478.	<i>Cora irene</i> Ris, 1918	23(22A+X)	—	Bolivia	Cumming 1964
479.	<i>Polythore boliviana</i> (McLachlan, 1878)	23(22A+X)	—	Bolivia	Cumming 1964
Euphaeidae					
480.	<i>Anisopleura comes</i> Hagen, 1880	25(24A+X)	+	Nepal	Kiauta and Kiauta 1976, 1982
481.	<i>Bayadera indica</i> (Selys, 1853)	25(24A+X)	+	Nepal	Chatterjee and Kiauta 1973
		— » —	+	Nepal	Kiauta 1975
482.	<i>Euphaea guerini</i> Rambur, 1842	25(24A+X)	—	Thailand	Kiauta and Kiauta 1983
483.	<i>Epallage fatime</i> (Charpentier, 1840)	25(24A+X)	—	Greece	Kiauta 1970b
		— » —	—	Greece	Chatterjee and Kiauta 1973
Megapodagrionidae					
484.	<i>Allopodagrion contortum</i> (Selys, 1862)	25(24A+X)	+	Brazil	Kiauta 1972b as <i>Megapodagrion contortum</i> (Selys, 1862)
485.	<i>Teinopodagrion macropus</i> (Selys, 1862)	25(24A+X)	—	Bolivia	Cumming 1964 as <i>Megapodagrion macropus</i> (Selys, 1862)
486.	<i>T. setigerum</i> (Selys, 1886)	25(24A+X)	—	Bolivia	Cumming 1964 as <i>Megapodagrion setigerum</i> Selys, 1886
Heteragrionidae					
487.	<i>Heteragrion flavidorsum</i> Calvert, 1909	25(24A+X)	—	Bolivia	Cumming 1964
488.	<i>H. inca</i> Calvert, 1909	25(24A+X)	+	Bolivia	Cumming 1964
Philogeniidae					
489.	<i>Philogenia carrillica</i> Calvert, 1907	25(24A+X)	+	Costa Rica	Cumming 1964
Hypolestidae					
490.	<i>Hypolestes clara</i> (Calvert, 1891)	l7(16A+X)	—	Jamaica	Cumming 1964
COENAGRIONOIDEA					
Platycnemididae					
491.	<i>Calicnemia miniata</i> (Selys, 1886)	25(24A+X)	+	Nepal	Kiauta and Kiauta 1982
492.	<i>C. pulverulans</i> (Selys, 1886)	25(24A+X)	—	Nepal	Kiauta 1975
493.	<i>Calicnemia</i> sp.	25(24A+X)	—	Nepal	Kiauta 1975
494.	<i>Calicnemia</i> sp.	25(24A+X)	—	India	Tyagi 1978b
495.	<i>Coeliccia chromothorax</i> (Selys, 1891)	25(24A+X)	—	India	Walia and Devi 2020b
496.	<i>C. bimaculata</i> (Laidlaw, 1914)	25(24A+X)	—	India	Walia and Devi 2020b
497.	<i>C. didyma</i> (Selys, 1863)	25(24A+X)	—	India	Walia and Devi 2020b
498.	<i>C. fraseri</i> (Laidlaw, 1932)	25(24A+X)	—	India	Walia and Devi 2020b
499.	<i>C. renifera</i> (Selys, 1886)	25(24A+X)	—	Nepal	Kiauta 1974, 1975
		— » —	—	India	Walia and Devi 2020b
500.	<i>Copera annulata</i> (Selys, 1863)	25(24A+X)	+	Japan	Kichijo 1941, 1942a, c
		— » —	+	India	Dasgupta 1957
		— » —	—	Thailand	Kiauta and Kiauta 1983
		— » —	+	India	Walia and Devi 2018

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
501.	<i>C. marginipes</i> (Rambur, 1842)	25(24A+X)	–	India	Tyagi 1978a, b
		– » –	–	Thailand	Kiauta and Kiauta 1983
		– » –	+	India	Walia and Devi 2018
502.	<i>C. vittata</i> (Selys, 1863)	25(24A+X)	+	India	Walia and Devi 2018
		– » –	+	India	Walia and Devi 2018 ( <i>C. v. assamensis</i> (Laidlaw, 1914))
503.	<i>Disparoneura quadrimaculata</i> (Rambur, 1842)	25(24A+X)	–	India	Walia and Devi 2020a
504.	<i>Esme cyaneovittata</i> Fraser, 1922	25(24A+X)	–	India	Walia and Devi 2020a
505.	<i>E. longistyla</i> Fraser, 1931	25(24A+X)	–	India	Walia and Devi 2020a
506.	<i>Onychargia atrocyana</i> (Selys, 1865)	25(24A+X)	–	Thailand	Kiauta and Kiauta 1983
507.	<i>Platycnemis pennipes</i> (Pallas, 1771)	25(24A+X)	–	Finland	Oksala 1945
		– » –	–	Italy	Kiauta 1971a
		– » –	–	Russia	Perepelov and Bugrov 2001b
508.	<i>Prodasineura autumnalis</i> (Fraser, 1922)	25(24A+X)	+	Thailand	Kiauta and Kiauta 1983
509.	<i>P. nigra</i> (Fraser, 1922)	25(24A+X)	–	India	Walia and Devi 2020a
510.	<i>P. verticalis</i> (Selys, 1860)	25(24A+X)	–	India	Walia and Devi 2020a
511.	<i>Prodasineura</i> sp.1	25(24A+X)	–	Thailand	Kiauta and Kiauta 1983
512.	<i>Prodasineura</i> sp.2	25(24A+X)	–	Thailand	Kiauta and Kiauta 1983
Coenagrionidae					
513.	<i>Acanthagrion ascendens</i> Calvert, 1909	27(26A+X)	+	Bolivia	Cumming 1964
514.	<i>A. chacoense</i> Calvert, 1909	27(26A+X)	+	Bolivia	Cumming 1964
515.	<i>A. gracile</i> (Rambur, 1842)	27(26A+X)	–	Surinam	Kiauta 1979a ( <i>A. g. minarum</i> Selys, 1876)
		– » –	–	Brazil	Ferreira et al. 1979 ( <i>A. g. minarum</i> Selys, 1876)
516.	<i>Aeolagrion</i> inca Selys, 1876	27(26A+X)	–	Bolivia	Cumming 1964 as <i>A. foliaceum</i> (Sjöstedt, 1918)
517.	<i>Agriocnemis clauseni</i> Fraser, 1922	27(26A+X)	+	India	Tyagi 1978a, b
518.	<i>A. femina</i> (Brauer, 1868)	27(26A+X)	–	Philippines	Kiauta and Kiauta 1980b
		– » –	+	Thailand	Kiauta and Kiauta 1983
519.	<i>A. pygmaea</i> (Rambur, 1842)	27(26A+X)	–	India	Tyagi 1978b
		– » –	+	Thailand	Kiauta and Kiauta 1983
520.	<i>Amphiagrion abbreviatum</i> (Selys, 1876)	27(26A+X)	–	USA	Cruden 1968
521.	<i>Amphiallagma parvum</i> (Selys, 1876)	27(26A+X)	+	India	Handa and Kochhar 1985 as <i>Enallagma parvum</i> Selys, 1876
522.	<i>Argia apicalis</i> (Say, 1839)	37(36A+X)	–	USA	Kiauta and Kiauta 1980b
523.	<i>A. fumipennis</i> (Burmeister, 1839)	27(26A+X)	–	USA	Kiauta and Kiauta 1980c ( <i>A. f. atra</i> Gloyd, 1968)
		– » –	–	USA	Kiauta and Brink 1978 ( <i>A. f. fumipennis</i> (Burmeister, 1839))
		– » –	–	USA	Kiauta and Kiauta 1980c ( <i>A. f. fumipennis</i> )
		– » –	+	Canada	Kiauta and Kiauta 1980c ( <i>A. f. violacea</i> (Hagen, 1861))
524.	<i>A. funebris</i> (Hagen, 1861)	27(26A+X)	–	USA	Kiauta 1972b
		28(26A+XX)*	–	Mexico	Kiauta and Kiauta 1980c
525.	<i>A. immunda</i> (Hagen, 1861)	27(26A+X)	–	USA	Kiauta and Kiauta 1980c
526.	<i>A. moesta</i> (Hagen, 1861)	25(24A+X)	–	Canada	Kiauta 1978
		– » –	–	USA	Kiauta and Kiauta 1980c
527.	<i>A. nahuana</i> Calvert, 1902	25(24A+X)	–	USA	Kiauta and Kiauta 1980c
528.	<i>A. sedula</i> (Hagen, 1861)	27(26A+X)	–	USA	Cruden 1968
		– » –	–	USA	Kiauta and Kiauta 1980c
529.	<i>A. tibialis</i> (Rambur, 1842)	37(36A+X)	–	USA	Kiauta and Kiauta 1980c
530.	<i>A. translata</i> Hagen, 1865	25(24A+X)	+	USA	Kiauta and Kiauta 1980c
531.	<i>A. violacea</i> (Hagen, 1861)	27(26A+X)	–	USA	Cruden 1968
532.	<i>A. vivida</i> (Hagen, 1861)	27(26A+X)	–	USA	Cruden 1968
533.	<i>Ceriagrion auranticum</i> Fraser, 1922	27(26A+X)	+	Thailand	Kiauta and Kiauta 1983 as <i>C. latericium</i> Liefstinck, 1951
534.	<i>C. azureum</i> (Selys, 1891)	27(26A+X)	–	Nepal	Kiauta 1974, 1975
535.	<i>C. cerinomelas</i> Liefstinck, 1927	27(26A+X)	–	Nepal	Kiauta 1974, 1975



Taxon	Karyotype formula 2n ♂	m-chromo somes	Country	References
536. <i>C. cerinorubellum</i> (Brauer, 1866)	27(26A+X)	+	India	Dasgupta 1957
	— » —	+	India	Prasad and Thomas 1992
537. <i>C. coromandelianum</i> (Fabricius, 1798)	27(26A+X)	+	India	Ray Chaudhuri and Dasgupta 1949
	— » —	+	India	Srivastava and Das 1953
	— » —	+	India	Das 1956
	— » —	+	Nepal	Kiauta and Kiauta 1982
	— » —	+	India	Prasad and Thomas 1992
538. <i>C. fallax</i> Ris, 1914	27(26A+X)	+	Republic of South Africa	Dasgupta 1957
539. <i>C. glabrum</i> (Burmeister, 1839)	27(26A+X)	—	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
540. <i>C. rubiae</i> Laidlaw, 1916	27(26A+X)	+	India	Asana and Makino 1935
	— » —	+	India	Makino 1935
	— » —	+	India	Kichijo 1942a
541. <i>C. tenellum</i> (Villers, 1789)	27(26A+X)	+	Italy	Kiauta 1971a ( <i>C. t. tenellum</i> (Villers, 1789))
542. <i>Chromagrion conditum</i> (Hagen, 1876)	27(26A+X)	—	USA	Cruden 1968
543. <i>Coenagrion armatum</i> (Charpentier, 1840)	27(26A+X)	—	Finland	Oksala 1939a
	— » —	—	Former USSR	Makalowskaja 1940
544. <i>C. hastulatum</i> (Charpentier, 1825)	27(26A+X)	—	Former USSR	Makalowskaja 1940
	— » —	—	Russia	Perepelov and Bugrov 2001b
545. <i>C. hylas</i> (Trybom, 1889)	27(26A+X)	—	Austria	Kiauta and Kiauta 1991 ( <i>C. h. freyi</i> (Bilek, 1954))
546. <i>C. lunulatum</i> (Charpentier, 1840)	27(26A+X)	+	Russia	Perepelov and Bugrov 2001b
547. <i>C. pulchellum</i> (Vander Linden, 1823)	27(26A+X)	—	Former USSR	Makalowskaja 1940
	— » —	—	Netherlands	Kiauta 1969c
	— » —	+	Russia	Kuznetsova et al. 2020b
548. <i>C. puella</i> (Linnaeus, 1758)	27(26A+X)	+	Russia	Kuznetsova et al. 2020b
549. <i>C. resolutum</i> (Hagen, 1876)	27(26A+X)	—	USA	Cruden 1968
550. <i>Coenagrion</i> sp.	27(26A+X)	+	Japan	Kichijo 1941, 1942d, e
551. <i>Diceratobasis macrogaster</i> (Selys, 1875)	27(26A+X)	+	Jamaica	Cumming 1964
552. <i>Enallagma aspersum</i> (Hagen, 1861)	27(26A+X)	—	USA	Cruden 1968
553. <i>E. boreale</i> Selys, 1875	27(26A+X)	—	USA	Cruden 1968
554. <i>E. carunculatum</i> Morse, 1895	27(26A+X)	—	USA	Cruden 1968
555. <i>E. circulatum</i> Selys, 1883	27(26A+X)	+	Russia	Perepelov and Bugrov 2001b
556. <i>E. civile</i> (Hagen, 1861)	27(26A+X)	—	USA	Cruden 1968
557. <i>E. cyathigerum</i> (Charpentier, 1840)	27(26A+X)	—	Finland	Oksala 1939a, 1945
	— » —	—	Former USSR	Makalowskaja 1940
	— » —	+	USA	Brink and Kiauta 1964
	27(26A+X), 29(28A+X)	—	USA	Cruden 1968
	27(26A+X) 29(28A+X)	+	Netherlands	Kiauta 1969a, c
558. <i>E. ebrium</i> (Hagen, 1861)	27(26A+X)	—	USA	Cruden 1968
559. <i>E. praevarum</i> (Hagen, 1861)	27(26A+X)	—	USA	Cruden 1968
560. <i>Erythromma lindeni</i> (Selys, 1840)	27(26A+X)	+	Italy	Kiauta 1971a
561. <i>E. najas</i> (Hansemann, 1823)	27(26A+X)	—	Finland	Oksala 1939a
	— » —	—	Former USSR	Makalowskaja 1940
	— » —	—	Netherlands	Kiauta 1969a
	— » —	—	Russia	Perepelov and Bugrov 2001b
	— » —	+	Russia	Kuznetsova et al. 2020b
562. <i>Homeoura chelifera</i> (Selys, 1876)	27(26A+X)	+	Surinam	Kiauta 1979a as <i>Enallagma cheliferum</i> (Selys, 1876)
	— » —	+	Brazil	Ferreira et al. 1979 as <i>E. cheliferum</i>
563. <i>Ischnura aurora</i> (Brauer, 1865)	27(26A+X)	—	Nepal	Kiauta 1974, 1975
	— » —	—	India	Handa and Kochhar 1985
564. <i>I. capreola</i> (Hagen, 1861)	27(26A+X)	—	Bolivia	Cumming 1964 as <i>Ceratura capreola</i> (Hagen, 1861)

Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
565.	<i>I. cervula</i> Selys, 1876	27(26A+X)	–	USA	Cruden 1968
566.	<i>I. denticollis</i> (Burmeister, 1839)	27(26A+X)	–	USA	Cruden 1968
567.	<i>I. elegans</i> (Van der Linden, 1823)	27(26A+X)	–	Finland	Oksala 1939a, 1945
		– » –	–	Netherlands	Kiauta 1969a
		– » –	–	Russia	Perepelov 2003
568.	<i>I. fluviatilis</i> Selys, 1876	27(26A+X)	–	Bolivia	Cumming 1964
569.	<i>I. forcipata</i> Morton, 1907	27(26A+X)	–	Nepal	Kiauta 1974, 1975
570.	<i>I. nursei</i> (Morton, 1907)	25(24A+X)	+	India	Tyagi 1978b as <i>Rhodischnura nursei</i> (Morton, 1907)
571.	<i>I. pumilio</i> (Charpentier, 1825)	27(26A+X)	+	Netherlands	Kiauta 1979b
572.	<i>I. perparva</i> Selys, 1876	27(26A+X)	–	USA	Cruden 1968
573.	<i>I. ramburii</i> (Selys, 1850)	27(26A+X)	+	USA	Kiauta and Brink 1978
574.	<i>I. rufostigma</i> Selys, 1876	27(26A+X)	–	Nepal	Kiauta 1974, 1975 ( <i>I. r. annandalei</i> Laidlaw, 1919)
575.	<i>I. senegalensis</i> (Rambur, 1842)	27(26A+X)	+	Japan	Kichijo 1941, 1942d, e
		– » –	+	India	Dasgupta 1957
		– » –	+	Ethiopia	Kiauta 1969b
		– » –	+	Philippines	Kiauta and Kiauta 1980b
		– » –	–	Thailand	Kiauta and Kiauta 1983
		– » –	+	India	Prasad and Thomas 1992
576.	<i>I. verticalis</i> (Say, 1839)	27(26A+X)	–	USA	Cruden 1968
577.	<i>I. ultima</i> Ris, 1908	27(26A+X)	–	Bolivia	Cumming 1964
578.	<i>Leptagrion macrurum</i> (Burmeister, 1839)	30(28A+neo-XY)	–	Brazil	Kiauta 1971c, 1972d
579.	<i>Mecistogaster</i> sp. 1	29(28A+X)	+	Bolivia	Cumming 1964
580.	<i>Mecistogaster</i> sp. 2	12(10A+neo-XY)	–	Bolivia	Cumming 1964
581.	<i>Megalagrion oahuense</i> (Blackburn, 1884)	27(26A+X)	+	Hawaii	Kiauta 1969b
582.	<i>Mortonagrion selenion</i> (Ris, 1916)	27(26A+X)	+	Japan	Kichijo 1941, 1942a, d, e
583.	<i>Nehalennia irene</i> (Hagen, 1861)	27(26A+X)	–	USA	Cruden 1968
584.	<i>N. speciosa</i> (Charpentier, 1840)	28(26A+XX)*	–	Finland	Oksala 1945
585.	<i>Oxyagrion hempeli</i> Calvert, 1909	27(26A+X)	–	Brazil	Souza Bueno 1982
586.	<i>O. terminale</i> Selys, 1876	27(26A+X)	–	Surinam	Kiauta 1979a
		– » –	–	Brazil	Ferreira et al. 1979
587.	<i>Paracercion hieroglyphicum</i> (Brauer, 1865)	27(26A+X)	+	Japan	Kichijo 1941, 1942d, e as <i>Coenagrion hieroglyphicum</i> (Brauer, 1865)
588.	<i>P. malayanum</i> (Selys, 1876)	27(26A+X)	+	Nepal	Kiauta 1974, 1975
589.	<i>Proischnura subfurcata</i> (Selys, 1876)	27(26A+X)	–	Kenya	Wasscher 1985 as <i>Enallagma subfurcatum</i> Selys, 1876
590.	<i>Pseudagrion acaciae</i> Förster, 1906	27(26A+X)	+	Republic of South Africa	Boyes et al. 1980
591.	<i>P. australasiae</i> Selys, 1876	27(26A+X)	+	India	Dasgupta 1957
592.	<i>P. decorum</i> (Rambur, 1842)	27(26A+X)	+	India	Dasgupta 1957
593.	<i>P. kersteni</i> (Gerstaker, 1869)	27(26A+X)	–	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
594.	<i>P. microcephalum</i> (Rambur, 1842)	27(26A+X)	+	India	Dasgupta 1957
		– » –	+	Philippines	Kiauta and Kiauta 1980b
595.	<i>P. pruinatum</i> (Burmeister, 1839)	27(26A+X)	+	Thailand	Kiauta and Kiauta 1983
596.	<i>P. rubripes</i> (Selys, 1876)	27(26A+X)	+	India	Dasgupta 1957
		– » –	+	Philippines	Kiauta and Kiauta 1980b
		– » –	+	Thailand	Kiauta and Kiauta 1983
597.	<i>P. salisburyense</i> Ris, 1921	27(26A+X)	+	Kingdom of Eswatini (Former Swaziland)	Boyes et al. 1980
598.	<i>P. spencei</i> Fraser, 1922	27(26A+X)	+	India	Dasgupta 1957
599.	<i>P. whellani</i> Pinhey, 1956	25(24A+X)	+	Burkina Faso (Former Voltaic Republic)	Kiauta and Ochssée 1979
600.	<i>Pyrrhosoma nymphula</i> (Sutzer, 1776)	28(26A+XX)*	–	Finland	Oksala 1945
601.	<i>Telebasis carmesina</i> Calvert, 1909	27(26A+X)	–	Surinam	Kiauta 1979a
		– » –	–	Brazil	Ferreira et al. 1979
602.	<i>Tigriagrion aurantinigrum</i> Calvert, 1909	27(26A+X)	–	Bolivia	Cumming 1964



Taxon		Karyotype formula 2n ♂	m-chromo somes	Country	References
603.	<i>Xanthocnemis zealandica</i> (McLachlan, 1873)	27(26A+X)	–	New Zealand	Jensen 1980 as <i>X. zelandica</i> (McLachlan, 1873)
604.	<i>Zoniagrion exclamationis</i> (Selys, 1876)	27(26A+X)	–	USA	Cruden 1968
<b>Protoneuridae</b>					
605.	<i>Caconeura autumnalis</i> Fraser, 1922	25(24A+X)	+	India	Tyagi 1978b
606.	<i>Epipleoneura</i> sp.	27(26A+X)	–	Bolivia	Cumming 1964
607.	<i>Protoneura rubriventris</i> (Selys, 1860)	27(26A+X)	+	Bolivia	Cumming 1964 as <i>Neoneura rubriventris</i> Selys, 1860

\* In the original publication, the female karyotype is given.

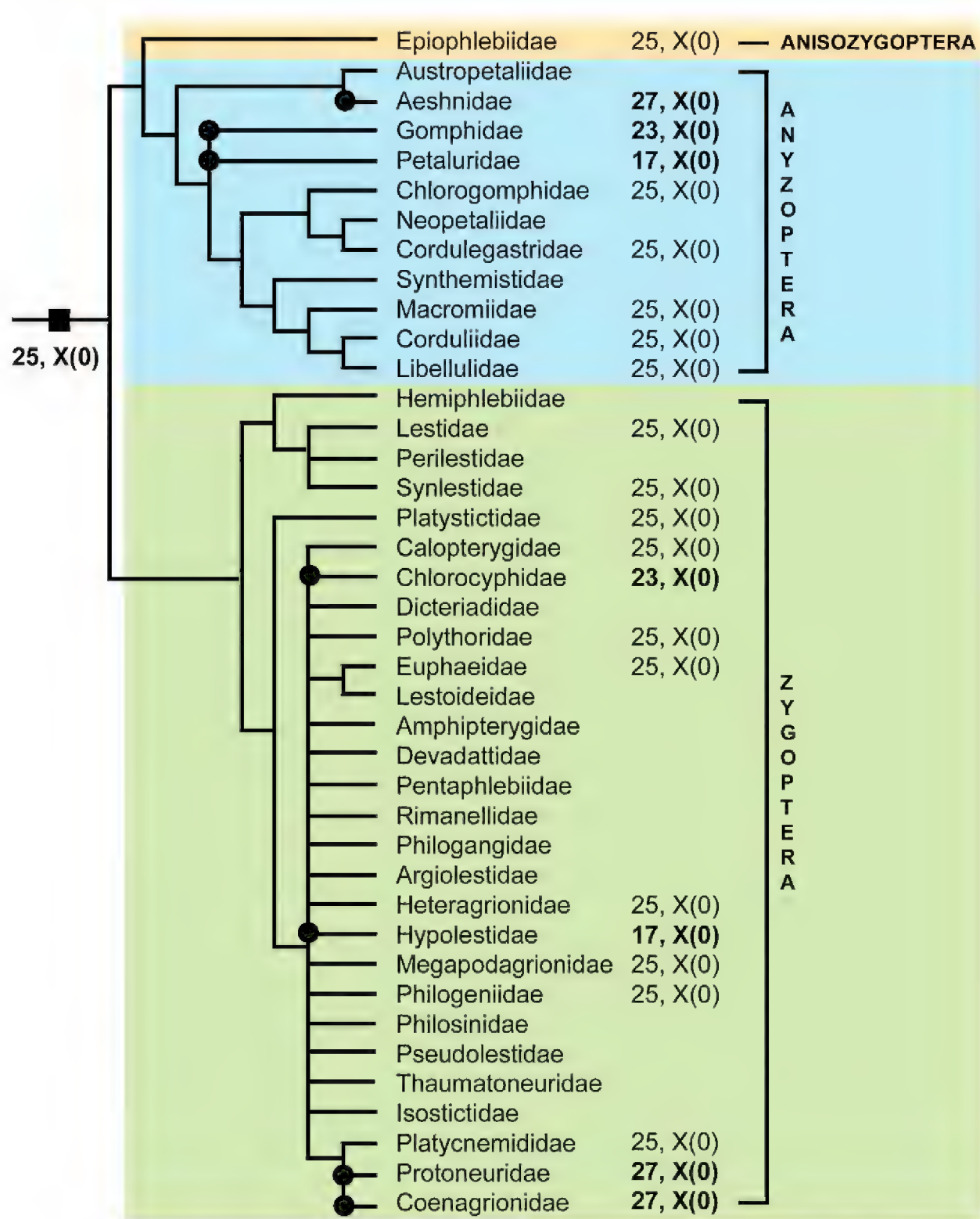
\*\* Jensen (1980) considers these data as erroneous (but see section “Concluding remarks and future directions” in the present paper).

\*\*\* Karyotype formula is extrapolated based on vague descriptions by Cumming (1964).

**Table 2.** The diversity of chromosome numbers and sex chromosome mechanisms, and modal karyotypes in 23 families of Odonata: a summary.

Taxa (N of species/genera described*)		N of species/ genera studied	Male karyotypes	Modal karyotype	N of species/genera with modal karyotype (occurrence in percent)
ANISOZYGOPTERA					
Epiophlebioidea	Epiophlebiidae (4/1)	1/1	25, X0	24A + X	1 (100) / 1 (100)
ANISOPTERA					
Aeshnoidea	Aeshnidae (456/51)	58/18	13, X0; 14, neo-XY; 15, X0; 16, neo-XY; 19, X0; 21, X0; 24, neo-XY; 25, X0; 26, neo-XY; 27, X0	26A + X	44 (76) / 14 (78)
Petaluroidea	Petaluridae (10/5)	4/3	17, X0; 19, X0; 25, X0	16A + X	3 (75) / 2 (67)
Gomphoidea	Gomphidae (980/87)	66/31	12, neo-neo-XY; 21, X0; 22, neo-XY; 23, X0; 24, neo-XY; 25, X0	22A + X	57 (86) / 28 (90)
Libelluloidea	Macromiidae (125/4)	6/3	25, X0	24A + X	6 (100) / 3 (100)
	Corduliidae (154/20)	23/7	10, neo-XY; 11, X0; 13, X0; 14, neo-XY; 20, XY; 21, X0; 25, X0; 26, neo-XY; 27, X0	24A + X	19 (83) / 6 (86)
	Libellulidae (1037/142)	255/59	6, neo-XY; 6 neo-XY; 8, neo-XY; 10, neo-XY; 12, neo-XY; 17, X0; 21, X0; 22, neo-XY; 23, X0; 23, X1X2Y; 24, neo-XY; 25, X0; 27, X0; 28, neo-XY; 29, X0; 41, X0	24A + X	227 (89) / 57 (97)
Cordulegastroidea	Cordulegastridae (46/3)	9/3	23, X0; 25, X0	24A + X	8 (89) / 3 (100)
	Chlorogomphidae (47/3)	1/1	25, X0	24A + X	1 (100) / 1 (100)
ZYGOPTERA					
Lestoidea	Lestidae (151/9)	20/5	19, X0; 21, X0; 25, X0	24A + X	18 (90) / 5 (100)
	Synlestidae (39/9)	1/1	25, X0	24A + X	1 (100) / 1 (100)
Platystictoidea	Platystictidae (224/6)	4/3	25, X0	24A + X	4 (100) / 3 (100)
Calopterygoidea	Calopterygidae (185/21)	20/8	23, X0; 25, X0; 27, X0	24A + X	20 (100) / 8 (100)
	Chlorocyphidae (144/19)	9/6	23, X0; 25, X0	22A + X	8 (89) / 5 (84)
	Polythoridae (59/7)	2/2	23, X0	22A + X	2 (100) / 2 (100)
	Euphaeidae (68/12)	4/4	25, X0	24A + X	4 (100) / 4 (100)
	Megapodagrionidae (296/42)	3/2	25, X0	24A + X	3 (100) / 2 (100)
	Heteragrionidae (57/2)	2/1	25, X0	24A + X	2 (100) / 1 (100)
	Philogeniidae (40/2)	1/1	25, X0	24A + X	1 (100) / 1 (100)
	Hypolestidae (6/4)	1/1	17, X0	16A + X	1 (100) / 1 (100)
	Platycnemididae (404/40)	22/8	25, X0	24A + X	19 (100) / 7 (100)
Coenagrionoidea	Coenagrionidae (1267/114)	92/28	12, neo-XY; 25, X0; 27, X0; 29, X0; 30, neo-XY; 37, X0	26A + X	81 (89) / 26 (90)
	Protoneuridae (260 / 25)	3/3	25, X0; 27, X0	26A + X	2 (70) / 2 (70)

\*Taken from Dijkstra et al. 2013



**Figure 1.** Mapping of modal karyotypes onto phylogenetic tree of Odonata families. The phylogenetic tree is taken from Bybee et al. (2016) who synthesized it based on trees from Dijkstra et al. (2014) and Carle et al. (2015). Plesiomorphic karyotype state is indicated by a black solid **square** (■), apomorphic karyotype states are indicated by black solid **circles** (●).

**Concluding remarks and future directions**

In total, karyotypes of 607 species (198 genera, 23 families) of Odonata are studied up to now. Table 1, presented in our work, includes 423 species (125 genera, 8 families) of the Anisoptera, 184 species (72 genera, 14 families) of the Zygoptera, and one species of the Anisozygoptera. Thus, the presently available karyotype data cover about 10% of the world species diversity of the order in general.



Within Odonata, chromosome numbers in males vary over a relatively wide range, from  $2n = 6$  in *Macrothemis hemichlora* and *Orthemis levis* to  $2n = 41$  in *O. nodiplaga*. Both low chromosome number species are suggested to have an evolutionarily secondary neo-XY system (Cumming 1964; Kiauta 1972c) that could have arisen through an X-autosome fusion from an X(0) system. All three of the above species belong to the largest dragonfly family Libellulidae, in which nearly 89% of studied species (255 in total) have the karyotype  $2n = 25(24A + X)$ . The last one is the most common in Odonata in general: it occurs in each of the three suborders, Zygoptera, Anisoptera and Anisozygoptera, and in all families with the exception of two damselfly families, the Polythoridae with only two studied species sharing  $2n = 23(22A + X)$  and a monotypic family Hypolestidae with  $2n = 17(16A + X)$  in male *Hypolestes clara*. Besides Libellulidae, the karyotype  $2n = 25(24A + X)$  is currently the presumed modal one in 14 other families, such being the case at least in six better covered (at species and/or generic level) families, i.e. the dragonfly families Corduliidae, Cordulegastridae, and Macromiidae, and the damselfly families Lestidae, Calopterygidae, and Platycnemididae (Table 2, Fig. 1). This chromosome set is suggested to be an ancestral one for the order Odonata in general (Oguma 1930; Kuznetsova et al. 2020b) although this suggestion remains questionable at this stage.

Chromosomal rearrangements, among which fission and fusions apparently predominated (Kiauta 1969c, 1972c), led to the appearance of divergent karyotypes in the evolution of Odonata. As a result, in many dragonfly and damselfly families, other karyotypes, when occurring, are of secondary origin as indicated by either a diverged number of autosomes or a secondary sex chromosome system of an XY-type or both (e.g. Cumming 1964; Kiauta 1969a, c; Agopian and Mola 1984, 1988; Mola et al. 1999; Perepelov and Bugrov 2002). Some interesting examples of this kind can be found in the family Libellulidae, in which  $2n = 25(24A + X)$  is most likely an evolutionarily initial karyotype (e.g. Agopian and Mola 1988). These examples are as follows (see Table 1): *Orthemis nodiplaga* and *O. ambinigra* with  $2n = 41(40A + X)$  and  $2n = 12(10A + \text{neo-XY})$ , respectively; *Erythrodiplax media* and *E. minuscula*, both with  $2n = 22(20A + \text{neo-XY})$ ; *Micrathyria longifasciata* and *M. unguolata* with  $2n = 24(22A + \text{neo-XY})$  and  $2n = 23(20A + X_1X_2Y)$ , respectively. In some families, any of these presumably derived karyotypes not only occurs but also prevails and may be considered modal (see Table 2 and Fig. 1). Within Anisoptera, such families are Aeshnidae ( $2n = 26A + X$ ) and Gomphidae ( $2n = 22A + X$ ), whereas within Zygoptera, these are Chlorocyphidae ( $2n = 22A + X$ ) and Coenagrionidae ( $2n = 26A + X$ ). Thus, Odonata, despite the fact that they have holokinetic chromosomes (Nokkala et al. 2002), demonstrate rather high karyotypic stability, with most species showing  $2n = 25$  (found in 60% of studied species),  $2n = 27$  (21%) and  $2n = 23$  (13%) which may point to some selective constraints acting to stabilize chromosome number in their evolution (Kuznetsova et al. 2020b).

There are the species for which different authors give various karyotypes that are sometimes difficult to interpret (see Table 1). In some cases, this might be due to

misidentifications of a particular species or an error in determining the karyotype. For example, Wolfe (1953) reported  $2n = 17(16A + X)$  for males of *Uropetala carovei* (Petaluridae, Anisoptera) from New Zealand. However, according to later studies of this species in the same locality (Jensen and Mahanty 1978; Jensen 1980), it has  $2n = 25(24A + X)$ , and Jensen (1980) therefore considers the Wolfe data as erroneous. We cannot exclude, however, that the above authors studied different *U. carovei* subspecies, *U. c. carovei* White, 1846 and *U. c. chiltoni* Tillyard, 1921, that may indeed have different karyotypes. In other cases, the chromosome number difference between geographic populations might be indicative of the inter-population variation within the bounds of one taxonomic species or even the existence of a species complex with several morphologically cryptic species. For example, 4 of the 17 studied species of the dragonfly genus *Aeshna* Fabricius, 1775 were reported to have different karyotypes in different populations. These are: *Aeshna grandis* –  $2n = 26A + X$  (former USSR),  $2n = 24A + X$  (former USSR, Finland), and  $2n = 24A + \text{neo-XY}$  (Netherlands, Finland); *A. isocles* –  $2n = 26A + X$  (USA) and  $2n = 24A + X$  (Russia); *A. juncea* –  $2n = 26A + X$  (Italy) and  $2n = 24A + \text{neo-XY}$  (Finland, former USSR, Italy); *A. mixta* –  $2n = 26A + X$  (Netherlands) and  $2n = 24A + X$  (India) (Table 1). In all such cases, special studies involving a combined analysis of karyotypes, morphology, distribution patterns and molecular markers are needed.

Approximately 80% of Odonata species have a pair of very small chromosomes, i.e. microchromosomes or m-chromosomes (Mola 2007, Table 1). A number of speculations have been forwarded to explain the origin of these chromosomes in Odonata. Kiauta (1968e) suggested m-chromosomes to be fragments of “normal” chromosomes, whereas Oguma (1930) considered them the remnants of an autosome pair in the process of its elimination by progressive loss of chromatin. The size of the smaller chromosome pair was shown to be variable within different species (Kiauta 1968e; see Mola 2007 for other references) which is consistent with both hypotheses. Closely related species and different populations of the same species often differ from each other in the presence/absence of m-chromosomes (Table 1). This is most likely due to the lack of clear criteria for the identification of a small chromosome pair as m-chromosomes in a particular karyotype (Mola 2007; Kuznetsova et al. 2020b).

Most cytogenetic studies of Odonata have been made only to determine the chromosome number and sex chromosome mechanism for which the routine staining was used. Although a considerable amount of such data was obtained (Table 1, 2), standard karyotypes of many Odonata taxa remain totally unknown (Fig. 1). Lack of data on more “primitive” families of Zygoptera (e.g. Hemiphlebiidae) and Anisoptera (e.g. Austropetaliidae and Neopetaliidae) makes difficult understanding karyotype evolution of the order in general.

During the last decades, karyotypes of a few dozen Odonata species were studied using various techniques of differential staining of chromosomes such as C-banding, AgNOR-staining and DNA specific fluorochrome banding visualizing constitutive heterochromatin, nucleolus organizing regions (NORs) and AT- and GC-rich chromosome segments, respectively. Such data can be found in the following publica-



tions: Thomas and Prasad (1986), Prasad and Thomas (1992), Perepelov et al. (1998), Perepelov and Bugrov (2001a, b, 2002), Grozeva and Marinov (2007), De Gennaro et al. (2008), Walia et al. (2011, 2018), Walia and Chahal (2014, 2018), Walia and Devi (2018), Walia and Katnoria (2018), Walia and Devi (2020a, b). Unfortunately, these data alone did not shed much light on the karyotypic evolution of Odonata.

Although the classical cytological techniques remain necessary starting points for cytogenetic studies of Odonata to get an overview of their genomes, the future of Odonata cytogenetics must be coupled with the application of new cytogenetic molecular techniques that enable the localization of specific DNA sequences in chromosomes and the identification of individual chromosomes in karyotypes. In the article by Frydrychová et al. (2004) and, on a larger scale, in two of our recent publications (Kuznetsova et al. 2018, 2020b), the fluorescence *in situ* hybridization (FISH) technique was used for the first time for analyzing Odonata karyotypes. Several species belonging to the Anisoptera (from the families Aeshnidae, Libellulidae, and Corduliidae) and the Zygoptera (from the families Coenagrionidae and Calopterygidae) were studied regarding the occurrence of the TTAGG telomeric repeats and the distribution of the *18S rRNA* genes in their karyotypes. The TTAGG repeats proved to be the canonical motif of telomeres in the class Insecta in general, which, however, was repeatedly lost in the evolution of different phylogenetic lineages (Kuznetsova et al. 2020a). It was shown in the listed Odonata publications that the (TTAGG)<sub>n</sub> motif does not occur in all but one (*Sympetrum vulgatum*) species, and the *18S* is located on one of the largest pairs of autosomes in all studied dragonfly species but on m-chromosomes in all studied damselfly species (Kuznetsova et al. 2020b).

The results obtained showed great promise of the combined use of FISH and classical and banding cytogenetics in order to identify new chromosomal markers, reveal differences between species, particularly when they share the same or very close karyotypes, and speculate about the mechanisms involved in the karyotype evolution of Odonata (Kuznetsova et al. 2020b). Another promising line of future research could be to test hypotheses (Mola and Papeschi 1994; Ardila-Garcia and Gregory 2009) about whether there is a relationship between karyotype evolution and genome size diversity in the Odonata or there is no such relationship.

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## References

- Agopian SS, Mola LM (1984) An exceptionally high chromosome number in *Orthemis nodipaga* Kersch (Anisoptera, Libellulidae). *Notulae Odonatologicae* 2(3): 45.
- Agopian SS, Mola LM (1988) Intra and interspecific karyotype variability in five species of Libellulidae (Odonata, Anisoptera). *Caryologia* 41(1): 69–78. <https://doi.org/10.1080/00087114.1988.10797849>
- Ardila-Garcia AM, Gregory TR (2009) An exploration of genome size diversity in dragonflies and damselflies (Insecta: Odonata). *Journal of Zoology* 278: 163–173. <https://doi.org/10.1111/j.1469-7998.2009.00557.x>
- Asana JJ, Makino S (1935) A comparative study of the chromosomes in the Indian dragonflies. *Journal of the Faculty of Science, Hokkaido University Series 6, Zoology* 4(2): 67–86.
- Boyes JW, van Brink JM, Kiauta B (1980) Sixteen dragonfly karyotypes from the republic of South Africa and Swaziland, with evidence on the possible hybrid nature of *Orthetrum julia falsum* Longfeild (Anisoptera: Libellulidae). *Odonatologica* 9: 131–145.
- Bybee S, Córdoba-Aguilar A, Duryea MC, Futahashi R, Hansson B, Lorenzo-Carballa MO, Schilder R, Stoks R, Suvorov A, Svensson EI, Swaegers J, Takahashi J, Watts PC, Wellenreuther M (2016) Odonata (dragonflies and damselflies) as a bridge between ecology and evolutionary genomics. *Frontiers in Zoology* 13: e46. <https://doi.org/10.1186/s12983-016-0176-7>
- Capitulo RA, Mola LM, Agopian SS (1991) Species catalogue and chromosomal data of Odonata from Argentina. *Revista de la Sociedad Entomológica Argentina* 49(1–4): 59–72.
- Carle FL, Kjer KM, May ML (2015) A molecular phylogeny and classification of Anisoptera (Odonata). *Arthropod Systematics and Phylogeny* 73(2): 281–301. <https://entomology.rutgers.edu/news/docs/Carle-2015-Anisoptera-Phylogeny-Classification.pdf>
- Carnoy BJ (1885) La cytodierese chez les arthropodes. IV. Pseudo-Nevropteres. *Cellule* 1: 279–282.
- Chatterjee K, Kiauta B (1973) Male germ cell chromosomes of two Calopterygoidea from the Darjeeling Himalaya (Zygoptera: Chlorocyphidae, Euphaeidae). *Odonatologica* 2(2): 105–108.
- Cruden RW (1968) Chromosome numbers of some North American dragonflies (Odonata). *Canadian Journal of Genetics and Cytology* 10: 200–214. <https://doi.org/10.1139/g68-029>
- Cumming RB (1964) Cytogenetic studies in the order Odonata. PhD thesis, University of Texas, Austin, 93 pp.
- Das C (1956) Studies on the association between non-homologous chromosomes during meiosis in four species of the Indian dragonflies (Odonata). *Journal of the Zoological Society of India* 8(2): 119–132.
- Dasgupta J (1957) Cytological studies of some Indian dragonflies. II: A study of the chromosomes during meiosis in thirty species of Indian Odonata (Insecta). *Proceedings of the Zoological Society of Calcutta* 10: 1–65.
- De Gennaro D (2004) Análisis meiótico y caracterización de la heterocromatina en especies argentinas de Anisoptera (Odonata). Tesis de Licenciatura. Facultad de Ciencias Exactas y Naturales de la Universidad de Buenos Aires. Buenos Aires, 66 pp.
- De Gennaro D, Rebagliati PJ, Mola LM (2008) Fluorescent banding and meiotic behaviour in *Erythrodiplax nigricans* (Libellulidae) and *Coryphaeschna perrensi* (Aeschnidae) (Anisoptera, Odonata). *Caryologia* 61: 60–67. <https://doi.org/10.1080/00087114.2008.10589610>



- Dijkstra K-DB, Bechly G, Bybee SM, Dow RA, Dumont HJ, Fleck G, Garrison RW, Hämäläinen M, Kalkman VJ, Karube H, May ML, Orr AG, Paulson DR, Rehn AC, Theischinger G, Trueman JWH, van Tol J, Ellenrieder N, Ware J (2013) The classification and diversity of dragonflies and damselflies (Odonata). In: Zhang ZQ (Ed.) Animal Biodiversity: An Outline of Higher-level Classification and Survey of Taxonomic Richness (Addenda 2013). Zootaxa 3703: 36–45. <https://doi.org/10.11646/zootaxa.3703.1.9>
- Dijkstra K-DB, Kalkman VJ, Dow RA, Stokvis FR, van Tol J (2014) Redefining the damselfly families: a comprehensive molecular phylogeny of Zygoptera (Odonata). Systematic Entomology 39: 68–96. <https://doi.org/10.1111/syen.12035>
- Ferreira A, Kiauta B, Zaha A (1979) Male germ cell chromosomes of thirty-two Brazilian dragonflies. Odonatologica 8: 5–22.
- Francović M, Jurečić R (1986) Prilog citogenetickim i citotaksonomskim istraživanjima vrste *Libellula depressa* L. (Odonata, Libellulidae). Plenarni Referati VII Kongres Biologa Jugoslavije, Budva, 341 pp.
- Francović M, Jurečić R (1989) Comparative cytogenetic analysis of karyotype morphology and organization in males of species *Libellula depressa* L. and *L. fulva* Müll. (Insecta: Odonata). Periodicum Biologorum 91(1): 32–33.
- Frydrychová R, Grossmann P, Trubač P, Vítková M, Marec F (2004) Phylogenetic distribution of TTAGG telomeric repeats in insects. Genome 47: 163–178. <https://doi.org/10.1139/g03-100>
- Fuchsówna J, Sawczyńska J (1928) Zachowanie się heterochromosomów podczas spermatogenezy u ważek (Odonata). Cz. I. *Aeschna grandis* L. *Libellula quadrimaculata* L. Archiwum Towarzystwa naukowego we Lwowie (III) 4(9): 177–197. [In Polish]
- Goni B, de Abenante YP (1982) Cytological notes on five dragonfly species from Uruguay. Odonatologica 11(4): 323–329.
- Grimaldi D, Engel MS (2005) Evolution of the Insects. Cambridge University Press, Cambridge, 755 pp.
- Grozeva SM, Marinov MG (2007) Cytogenetic study of *Somatochlora borisi* Marinov, 2001 (Odonata: Corduliidae), and three relative species. Acta Zoologica Bulgarica 59(1): 53–58.
- Handa SM, Batra HN (1980) Cytology of ten species of dragonflies (Anisoptera: Odonata). Proceedings of the 67<sup>th</sup> Indian Science Congress, Part III, Calcutta, 103 pp.
- Handa SM, Kochhar N (1985) Chromosomal architecture in two species of damselflies from Chandigarh and its surrounding areas. National Seminar on Current Trends in Chromosome Dynamics, Chandigarh, 34 pp.
- Handa SM, Mittal OP, Batra HN (1984) Chromosomes in ten species of dragonflies (Anisoptera: Odonata). Research Bulletin of the Panjab University (Science) 35: 65–75.
- Higashi K, Kayano H (1993) The distribution of distinct karyomorphs of *Crocothemis servilia* Drury (Anisoptera, Libellulidae) in Kyushu and the south-western islands of Japan. Japanese Journal of Entomology 61: 1–10.
- Higashi K, Lee CE, Kayano H, Kayano A (2001) Korea strait delimiting distribution of distinct karyomorphs of *Crocothemis servilia* (Drury) (Anisoptera: Libellulidae). Odonatologica 30(3): 265–270.
- Hirai H (1956) Chromosomes of six species of dragonflies. Zoological Magazine, Tokyo 65: 198–202.

- Hogben L (1921) Studies on synapsis, III. The nuclear organisation of the germ cells in *Libellula depressa*. Philosophical Transactions of the Royal Society of London Series B 92: 60–80. <https://doi.org/10.1098/rspb.1921.0006>
- Hung ACF (1971) Cytological studies of five dragonflies (Odonata: Anisoptera). Entomological News 82: 103–106.
- Jensen AL (1980) The karyotypes of five species of Odonata endemic to New Zealand. Odonatologica 9: 29–33.
- Jensen AL, Mahanty HK (1978) A preliminary note on the chromosome number of *Uropetala carovei* (White) (Anisoptera: Petaluridae). Odonatologica 7: 385–386.
- Kalkman VJ, Clausnitzer V, Dijkstra K-DB, Orr AG, Paulson DR, van Tol J (2008) Global diversity of dragonflies (Odonata) in freshwater. Hydrobiologia 595: 351–363. <https://doi.org/10.1007/s10750-007-9029-x>
- Katatani N (1987) On the chromosomes of dragonflies, 1. Synopsis on the studies in some Japanese dragonflies. Aeschna 20: 21–31.
- Kiauta B (1965) The chromosome behaviour in spermatogenetic meiosis of *Anax imperator* Leach (Odonata: Aeshnidae). Tombo 7(3–4): 18–21.
- Kiauta B (1966) The chromosome behaviour in spermatogenetic meiosis of the dragonfly *Sympetrum striolatum* (Charp.) (Odonata: Libellulidae) from Luxembourg. Bulletin de la Société des Naturalistes Luxembourgeois 69: 54–60.
- Kiauta B (1967a) Evolution of the chromosome complement in Odonata. Genen en Phaenen 11(4): 56–61.
- Kiauta B (1967b) Abstract. Evolution of the chromosome complement in Odonata. Genetica 38(3): 403–404. <https://doi.org/10.1007/BF01507474>
- Kiauta B (1967c) A new hypothesis on the evolution of the chromosome complement in Odonata. Tombo 10(1–4): 29–33.
- Kiauta B (1967d) Considerations on the evolution of the chromosome complement in Odonata. Genetica 38(4): 430–446. <https://doi.org/10.1007/BF01507474>
- Kiauta B (1967e) Meiotic chromosome behaviour in the male damselfly, *Calopteryx virgo* (Linnaeus), with a discussion on the value of chromosome numbers and karyotype morphology in odonate systematics. Deutsche Entomologische Zeitschrift 14(3–4): 339–348. <https://doi.org/10.1002/mmnd.19670140312>
- Kiauta B (1968a) Evolution of the chromosome complement in Odonata. Entomologische Berichten, Amsterdam 28(5): 97–100.
- Kiauta B (1968b) Morphology and kinetic behaviour of the odonate sex chromosomes, with a review of the distribution of sex determining mechanisms in the order. Genen en Phaenen 12(1): 21–24.
- Kiauta B (1968c) The chromosome numbers of eight Old World dragonflies (Odonata). Chromosome Information Service, Tokyo 9: 3–4.
- Kiauta B (1968d) The chromosomes of the male dragonfly *Cordulegaster boltoni* (Donovan, 1807) (Odonata: Cordulegasteridae). Biološki Vestnik: glasilo slovenskih biologov 16: 87–94.
- Kiauta B (1968e) Variation in size of the m-chromosome of the dragonfly, *Calopteryx virgo* (L.), and its significance for the chorogeography and taxonomy of the *Calopteryx virgo superspecies*. Genen en Phaenen 12(1): 11–16.

- Kiauta B (1968f) Variation in size of the dragonfly m-chromosome, with considerations on its significance for the chorogeography and taxonomy of the order Odonata, and notes on the validity of the rule of Reinig. *Genetica* 39(1): 64–74. <https://doi.org/10.1007/BF02324456>
- Kiauta B (1969a) Sex chromosomes and sex determining mechanisms in Odonata, with a review of the cytological conditions in the family Gomphidae, and reference to the karyotypic evolution in the order. *Genetica* 40(2): 127–157. <https://doi.org/10.1007/BF01787346>
- Kiauta B (1969b) The chromosomes of eight dragonfly species from continental Africa and Madagascar (Odonata). *Arnoldia* (Rhodesia) 4(15): 1–8.
- Kiauta B (1969c) Autosomal fragmentations and fusions in Odonata and their evolutionary implications. *Genetica* 40(2): 158–180. <https://doi.org/10.1007/BF01787347>
- Kiauta B (1969d) The chromosomes of the Hawaiian endemic dragonflies, *Megalagrion oahuense* (Blackburn) (Coenagrionidae: Pseudagrioninae) and *Nesogonia blackburni* (McLachlan) (Libellulidae: Sympetrinae), with a note on the cytotaxonomic affinities between the genera *Nesogonia* Kirby and *Sympetrum* Newman (order Odonata). *Proceedings of the Hawaiian Entomological Society* 20(2): 429–433.
- Kiauta B (1970a) The chromosomes of four Neotropical dragonflies from Mexico. *Chromosome Information Service*, Tokyo 11: 8–9.
- Kiauta B (1970b) The karyotype of the damselfly, *Epallage fatime* (Charpentier, 1840) (Odonata, Zygoptera: Epallagidae), with a note on the cytotaxonomic affinities in the superfamily Calopterygoidea. *Genetica* 41: 390–397. <https://doi.org/10.1007/BF00958931>
- Kiauta B (1971a) Studies on the germ cell chromosome cytology of some cytotaxonomically interesting or hitherto not studied Odonata from the autonomous region Friuli-Venezia Giulia (northern Italy). *Atti del Museo civico di Storia naturale di Trieste* 27: 65–127.
- Kiauta B (1971b) An unusual case of precocious segregation and chromosome fragmentation in the primary spermatocytes of the damselfly, *Calopteryx virgo meridionalis* (Selys, 1873), as evidence for a possible hybrid character of some populations of the *Calopteryx-virgo*-complex (Odonata, Zygoptera: Calopterygidae). *Genen en Phaenen* 14(2): 32–40.
- Kiauta B (1971c) Cytotaxonomic peculiarities in the neotropical odonate genera *Leptagrion* Selys, *Orthemis* Hagen and *Macrothemis* Hagen. Abstracts of papers read at the 1<sup>st</sup> European Symposium on Odonatology, Gent, 27–28.
- Kiauta B (1972a) Notes on new or little known dragonfly karyotypes, 2. Male germ cell chromosomes of four East Mediterranean species: *Lestes barbarus* (Fabricius), *Calopteryx splendens amasina* Bartenev (Zygoptera: Lestidae, Calopterygidae), *Caliaeschna microstigma* (Schneider) and *Orthetrum taeniolatum* (Schneider) (Anisoptera: Aeshnidae, Libellulidae). *Genen en Phaenen* 15: 95–98.
- Kiauta B (1972b) Notes on new or little known dragonfly karyotypes, 1. The germ cell chromosomes of three Latin American species: *Argia funebris* (Hagen), *Megapodagrion contortum* (Selys) (Zygoptera: Coenagrionidae, Megapodagrionidae) and *Castoraeschna castor* (Brauer) (Anisoptera: Aeshnidae). *Genen en Phaenen* 15: 23–26.
- Kiauta B (1972c) Synopsis on the main cytotaxonomic data in the order Odonata. *Odonatologica* 1(2): 73–102.
- Kiauta B (1972d) The karyotype of the damselfly, *Leptagrion macrurum* (Burmeister, 1839), and its possible origin, with a note on the cytotaxonomic affinities of the genus (Zygoptera: Coenagrionidae). *Odonatologica* 1(1): 31–35.



- Kiauta B (1973a) Notes on new or little known dragonfly karyotypes. III. Spermatocyte chromosomes of four Nearctic anisopterans: *Aeshna californica* Calvert (Aeshnidae), *Cordulia shurtleffi* Scudder (Corduliidae), *Sympetrum internum* Montgomery, and *S. madidum* (Hagen) (Libellulidae). *Genen en Phaenen* 16(1): 7–12.
- Kiauta B (1973b) Notes on new or little known dragonfly karyotypes. IV. Spermatocyte chromosomes of *Calopteryx splendens splendens* Harris (Zygoptera: Calopterygidae), *Gomphus pulchellus* Selys, *Libellula depressa* Linnaeus (Anisoptera: Gomphidae, Libellulidae) from northern France. *Genen en Phaenen* 16(2): 55–60.
- Kiauta B (1974) Introduction to insect cytotaxonomy. Lectures delivered at the Tribhuvan University, Kathmandu, Vol. 1. Nepal Research Center, Kathmandu, 81 pp.
- Kiauta B (1975) Cytotaxonomy of dragonflies, with special reference to the Nepalese fauna. Lectures delivered at the Tribhuvan University, Kathmandu, Vol. 2. Nepal Research Center, Kathmandu, 78 pp.
- Kiauta B (1977) Notes on new or little known dragonfly karyotypes.V. The male germ cell chromosomes of *Macromia moorei* Selys from Nepal (Anisoptera: Corduliidae, Epophthalmiinae). *Genen en Phaenen* 19: 49–51.
- Kiauta B (1978) Two cytotaxonomically interesting cases of irreversible autosome fusion in dragonflies *Agria modesta* (Hagen) (Zygoptera: Coenagrionidae) and *Anaciaeschna isosceles* (Müller) (Anisoptera: Aeshnidae). *Notulae Odonatologicae* 1(1): 7–9.
- Kiauta B (1979a) The karyotypes of some Anisoptera from Surinam. *Odonatologica* 2: 267–283.
- Kiauta B (1979b) The karyotype of *Ischnura pumilio* (Charp.) (Zygoptera: Coenagrionidae). *Notulae Odonatologicae* 1(3): 47–48.
- Kiauta B (1983) The status of the Japanese *Crocothemis servilia* (Drury) as revealed by karyotypic morphology (Anisoptera: Libellulidae). *Odonatologica* 12: 381–388.
- Kiauta B, Boyes JW (1972) Cytology of ten South American Libellulidae, with cytophylogenetic consideration of the genera *Orthemis* Hagen and *Erythrodipax* Brauer (Odonata, Anisoptera). *Genetica* 43(3): 407–421. <https://doi.org/10.1007/BF00156136>
- Kiauta B, Brink JM (1975) Cytotaxonomic notes on the *Sympetrum pedemontanum* complex (Anisoptera: Libellulidae). *Odonatologica* 4(4): 249–254.
- Kiauta B, Brink JM (1978) Male chromosome complements of some Florida dragonflies, United States. *Odonatologica* 7(1): 155–25.
- Kiauta B, Kiauta MAJE (1976) The chromosomes of some dragonflies from the Langtang Valley, Central Nepal. *Odonatologica* 5(4): 347–354.
- Kiauta B, Kiauta MAJE (1979) The karyotype of *Libellula fulva* Müll, from Switzerland (Anisoptera: Libellulidae). *Notulae Odonatologicae* 1(4): 73–74.
- Kiauta B, Kiauta MAJE (1980a) The karyotypes of *Aeshna subarctica elisabethae* Djak. and *Somatochlora alpestris* (Sel.) from Switzerland (Anisoptera, Aeshnidae, Corduliidae). *Notulae Odonatologicae* 1(6): 104–105.
- Kiauta B, Kiauta MAJE (1980b) On a small collection of dragonfly karyotypes from the Philippines. *Odonatologica* 9(3): 237–245.
- Kiauta B, Kiauta MAJE (1980c) Introduction to the cytotaxonomy of the odonate genus *Agria* Rambur (Zygoptera: Coenagrionidae). *Odonatologica* 9(1): 35–56.
- Kiauta B, Kiauta MAJE (1982) The chromosome numbers of sixteen dragonfly species from the Arun Valley, Eastern Nepal. *Notulae Odonatologicae* 9(1): 143–146.

- Kiauta B, Kiauta MAJE (1983) The chromosome numbers of some Odonata from Thailand. *Notulae Odonatologicae* 2(2): 17–32.
- Kiauta B, Kiauta MAJE (1991) Biogeographic considerations on *Coenagrion hylas freyi* (Bilek, 1954), based mainly on the karyotype features of a population from North Tyrol, Austria (Zygoptera: Coenagrionidae). *Odonatologica* 20(4): 417–431.
- Kiauta B, Kiauta MAJE (1995) The karyotypes of *Somatochlora meridionalis* Nielsen from Slovenia and *S. metallica* (Vander L.) from Switzerland, with a tentative note on the origin of Central European *S. metallica* (Odonata: Corduliidae). *Opuscula zoologica fluminensia* 137: 1–5.
- Kiauta B, Kiauta-Brink MAJE (1975) Chromosomes of the dragonfly, *Sympecma annulata braueri* (Yakobson & Bianki, 1905) from the Netherlands, with a note on the classification of the family Lestidae (Odonata, Zygoptera). *Genen en Phaenen* 18(2–3): 39–48.
- Kiauta B, Ochssée BV (1979) Some dragonfly karyotypes from the Voltiac Republic (Haute Volta), West Africa. *Odonatologica* 8: 47–54.
- Kichijo H (1939) Chromosomes of *Tachopteryx pryeri* and *Gomphus hakiensis* (Odonata, Aeshnidae). *Japanese Journal of Genetics* 15: 287–289. <https://doi.org/10.1266/jjg.15.287>
- Kichijo H (1941) Chromosomes of seven species of insects belonging to the order of dragonflies, suborder of damselflies. *Nagasaki Medical Journal* 19(10): 2033–2041. [In Japanese]
- Kichijo H (1942a) Insect chromosomes. IV. Order of dragonflies, Pt. 2. *Nagasaki Medical Journal* 20(10): 1639–1648. [In Japanese]
- Kichijo H (1942b) Insect chromosomes. III. Order of dragonflies, Pt. 1. *Nagasaki Medical Journal* 20(7): 1084–1092. [In Japanese]
- Kichijo H (1942c) Chromosomes of *Sympetrum eroticum eroticum* (Odonata). *Japanese Journal of Genetics* 18: 195–196. <https://doi.org/10.1266/jjg.18.195>
- Kichijo H (1942d) A comparative study of seven species of Zygoptera from Japan. *Acta medica Nagasakiensi* 3(2): 95–97.
- Kichijo H (1942e) On the chromosomes of some species of the zygopterous dragonflies (Odonata, Zygoptera). *Japanese Journal of Genetics* 18: 273–276. <https://doi.org/10.1266/jjg.18.273>
- Kumari U, Gautam DC (2017) Karyotypic studies on two species of *Orthetrum* (Anisoptera: Odonata) from Himachal Pradesh. *The Journal of Cytology and Genetics* 18: 1–7.
- Kuznetsova V, Grozeva S, Gokhman V (2020a) Telomere structure in insects: A review. *Journal of Zoological Systematics and Evolutionary Research* 58: 127–158. <https://doi.org/10.1111/jzs.12332>
- Kuznetsova VG, Maryńska-Nadachowska A, Shapoval NA, Anokhin BA, Shapoval AP (2018) Cytogenetic characterization of eight Odonata species originating from the Curonian Spit (the Baltic Sea, Russia) using C-banding and FISH with 18S rDNA and telomeric (TTAGG)<sub>n</sub> probes. *Cytogenetic and Genome Research* 153: 147–157. <https://doi.org/10.1159/000486088>
- Kuznetsova VG, Maryńska-Nadachowska A, Anokhin BA, Shapoval NA, Shapoval AP (2020b) Chromosomal analysis of eight species of dragonflies (Anisoptera) and damselflies (Zygoptera) using conventional cytogenetics and FISH: insights into the karyotype evolution of the ancient insect order Odonata. *Journal of Zoological Systematics and Evolutionary Research* 58, 00: 1–13. <https://doi.org/10.1111/jzs.12429> [in press]
- Lefevre G, McGill C (1908) The chromosomes of *Anasa tristis* and *Anax junius*. *The American Journal of Anatomy* 7(4): 469–487. <https://doi.org/10.1002/aja.1000070404>

- Makalowskaja WN (1940) Comparative karyological studies of dragonflies (Odonata). Archives russes d'Anatomie, d'Histologie et d'Embryologie 25: 24–39.
- Makino S (1935) A comparative study of the chromosomes in the Indian dragonflies. Japanese Journal of Genetics 11: 234–235. <https://doi.org/10.1266/jjg.11.234>
- McGill C (1904) The spermatogenesis of *Anax junius*. University of Missouri Studies 2: 236–250.
- McGill C (1907) The behavior of the nucleoli during oogenesis of the dragonfly with special reference to synapsis. Zoologische Jahrbücher. Abteilung für Anatomie und Ontogenie der Tiere 23: 207–230.
- Mola LM (1995) Post-reductional meiosis in *Aeshna* (Aeshnidae, Odonata). Hereditas 122: 47–55. <https://doi.org/10.1111/j.1601-5223.1995.00047.x>
- Mola LM (1996) Meiotic studies in nine species of *Erythrodiplax* (Libellulidae, Odonata). Neo-XY sex chromosome system in *Erythrodiplax media*. Cytologia 61: 349–357. <https://doi.org/10.1508/cytologia.61.349>
- Mola LM (2007) Cytogenetics of American Odonata. In: Tyagi BK (Ed.) Odonata: Biology of Dragonflies. Scientific Publishers, India, 153–173.
- Mola LM, Agopian SS (1985) Observations on the chromosomes of four South American Libellulidae (Anisoptera). Odonatologica 14(2): 115–125.
- Mola LM, Papeschi AG (1994) Karyotype evolution in *Aeshna* (Aeshnidae: Odonata). Hereditas 121: 185–189. <https://doi.org/10.1111/j.1601-5223.1994.00185.x>
- Mola LM, Papeschi AG, Carrillo ET (1999) Cytogenetics of seven species of dragonflies. Hereditas 131: 147–153. <https://doi.org/10.1111/j.1601-5223.1999.00147.x>
- Nokkala S, Laukkanen A, Nokkala C (2002) Mitotic and meiotic chromosomes in *Somatochlora metallica* (Corduliidae, Odonata). The absence of localized centromeres and inverted meiosis. Hereditas 136: 7–12. <https://doi.org/10.1034/j.1601-5223.2002.1360102.x>
- Oguma K (1915) A study of the chromosomes of dragonflies. Zoological Magazine 27: 241–250. [In Japanese]
- Oguma K (1917) Entomology and cytology. In: Nagano K (Ed.) A Collection of Essays for Mr. Yasushi Nawa, Written in Commemoration of His Sixtieth Birthday, October 8, 1917. Gifu, 105–114.
- Oguma K (1930) A comparative study of the spermatocyte chromosome in allied species of the dragonfly. Journal of Faculty of Sciences, Hokkaido University VI: 1–32.
- Oguma K (1942) Observationes de formis compositionibusque chromosomatum et dispositionibus eorum in tempore divisionis atque propositio aliquorum novorum terminorum. Japanese Journal of Genetics 18: 205–216. <https://doi.org/10.1266/jjg.18.205>
- Oguma K (1951) The chromosomes of *Epiophlebia superstes* Selys (dragonfly). Iden-no-Sogo-Kenkyu 2: 23–26.
- Oguma K, Asana JJ (1932) Additional data to our knowledge on the dragonfly chromosome with a note on the occurrence of X-Y chromosome in the ant-lion (Neuroptera). Journal of Faculty of Sciences, Hokkaido University 1(4): 133–142.
- Oksala T (1939a) Über Tetraploidie der Binde- und Fettgewebe bei den Odonaten. Hereditas 25: 132–144. <https://doi.org/10.1111/j.1601-5223.1939.tb02690.x>
- Oksala T (1939b) Über die somatische Polyploidie bei Insekten. Annales Entomologici Fennici 5(3): 208–218.



- Oksala T (1943) Zytologische Studien an Odonaten I. Chromosomenverhältnisse bei der Gattung *Aeschna* mit besonderer Berücksichtigung der postreduktionellen Teilung der Bivalente. *Annales Academiae Scientiarum Fennicae (A) IV, Biologica* (4): 1–64.
- Oksala T (1944) Zytologische Studien an Odonaten. II. Die Entstehung der Meiotischen Präkozytät. *Annales Academiae Scientiarum Fennicae (A) IV, Biologica* (5): 1–33.
- Oksala T (1945) Zytologische Studien an Odonaten. III. Die Ovogenese. *Annales Academiae Scientiarum Fennicae (A) IV, Biologica* (9): 1–132.
- Oksala T (1952) Chiasma formation and chiasma interference in the Odonata. *Hereditas* 38: 449–480. <https://doi.org/10.1111/j.1601-5223.1952.tb02937.x>
- Omura T (1949) On at-random connection of chromosomes in the aeschnid dragonfly, *Ictinus rapax*. *Japanese Journal of Genetics* 24: 162–165. <https://doi.org/10.1266/jjg.24.162>
- Omura T (1952) The spermatogenesis of an Indian dragonfly, *Ictinus rapax* (Rambur) with special reference to the behaviour of the spermatozoa in the cyst. *Biological Journal of Okayama University* 1(1–2): 103–146.
- Omura T (1953) On the abnormal spermatogenesis in an Indian dragonfly, *Ictinus rapax* (Rambur). *Biological Journal of Okayama University* 1(3): 163–170.
- Omura T (1955) A comparative study of the spermatogenesis in the Japanese dragonflies. I. Family Libellulidae. *Biological Journal of Okayama University* 2(2–3): 95–135.
- Omura T (1957) A comparative study of the spermatogenesis in the Japanese dragonfly II: Family Aeschnidae, Gomphidae and Calopterygidae. *Biological Journal of Okayama University* 3: 1–86.
- Perepelov EA (2003) Karyotype evolution of Odonata (Insecta) of Northern Palearctics. Ph.D. Dissertation, Novosibirsk, Russian Federation: Institute of Systematics and Ecology of Animals of Siberian Branch of Russian Academy of Sciences, 144 pp. [In Russian] <https://www.dissercat.com/content/evolyutsiya-kariotipov-strekoz-insecta-odonata-severnoi-palearktiki>
- Perepelov E, Bugrov AG (2001a) C-heterochromatin in chromosomes of *Ophiogomphus Cecilia cecilia* (Four.) (Anisoptera: Gomphidae) with notes on the sex chromosome origin in the species. *Caryologia* 54(2): 169–172. <https://doi.org/10.1080/00087114.2001.10589224>
- Perepelov E, Bugrov AG (2001b) The constituent heterochromatin in karyotypes of dragonflies. *Belyshevia* 1(1): 10–13. [In Russian]
- Perepelov E, Bugrov AG (2002) Constitutive heterochromatin in chromosomes of some Aeshnidae, with notes on the formation of the neo-XY/neo-XX mode of sex determination in *Aeshna* (Anisoptera). *Odonatologica* 31(1): 77–83.
- Perepelov EA, Bugrov AG, Warchałowska-Śliwa E (1998) C banded karyotypes of some dragonfly species from Russia. *Folia biologica (Kraków)* 46: 137–142.
- Perepelov EA, Bugrov AG, Warchałowska-Śliwa E (2001) C-banded karyotypes of some dragonfly species from Russia. II. The families Cordulegasteridae, Corduliidae and Gomphidae. *Folia biologica (Kraków)* 49(3–4): 175–178.
- Prasad K, Thomas KI (1992) C-band pattern homogeneity in dragonflies (Odonata). *Caryologia* 45: 57–68. <https://doi.org/10.1080/00087114.1992.10797211>
- Ray Chaudhuri SP, Dasgupta J (1949) Cytological studies on the Indian dragonflies I. Structure and behaviour of chromosomes in six species of dragonflies (Odonata). *Proceedings of the Zoological Society of Bengal* 2: 81–93.



- Rehn AC (2003) Phylogenetic analysis of higher-level relationships of Odonata. *Systematic Entomology* 28: 181–240. <https://doi.org/10.1046/j.1365-3113.2003.00210.x>
- Sandhu R, Malhotra I (1994a) Karyological studies of four aeshnid dragonflies from the states of Jammu and Kashmir and Himachal Pradesh (India). In: Srivastava VK (Ed.) *Advances in Oriental Odonatology: Proceedings of IV South Asian Symposium of Odonatology*, Allahabad, India October 10–12, 1992, Cherry publications, Allahabad, 111–115.
- Sandhu R, Malhotra I (1994b) New chromosome count in male dragonfly, *Anatogaster s. basalis*. *Bionature* 14: 69–70.
- Sandhu R, Walia GK (1995) A note on the karyotype of *Potamarcha congener* (Anisoptera: Libellulidae). *Chromosome Information Service* 58: 24–25.
- Sangal SK, Tyagi BK (1982) The spermatocyte chromosomes of *Anax immaculifrons* Rambur from India (Anisoptera: Aeshnidae). *Notulae odonatologicae* 1(9): 154–155.
- Schorr M, Paulson D (2020) World Odonata List. <https://www.pugetsound.edu/academics/academic-resources/slater-museum/biodiversity-resources/dragonflies/world-odonata-list2/>
- Seshachar BR, Bagga S (1962) Chromosome number and sex-determining mechanism in dragonfly *Hemianax ephippiger* (Burmeister). *Cytologia* 27: 443–449. <https://doi.org/10.1508/cytologia.27.443>
- Seshachar BR, Bagga S (1963) A cytochemical study of oogenesis in the dragonfly *Pantala flavescens* (Fabricius). *Growth* 27: 225–246.
- Sharma OP, Durani S (1995) A study on the chromosomes of three species of dragonflies (Odonata: Anisoptera). *National Academy Science Letters* 18(5–6): 97.
- Smith EA (1916) Spermatogenesis of the dragonfly *Sympetrum semicinctum* with remarks upon *Libellula basalis*. *Biological Bulletin* 31: 269–290. <https://doi.org/10.2307/1536236>
- Souza Bueno AM (1982) Estudos cromossomicos na ordem Odonata. M. Sc. Thesis, Universidade Estadual Paulista, 140 pp.
- Srivastava MDL, Das CC (1953) Heteropycnosis in the autosome segments of *Ceriagrion coromandelianum* (Odonata). *Nature* 172: 765–766. <https://doi.org/10.1038/172765b0>
- Suzuki KJ, Saitoh K (1990) A revised chromosome study of Japanese Odonates (I). Chromosomes of 14 species belonging to nine families. *The Science Reports of the Hirosaki University* 37: 38–49.
- Suzuki KJ, Saitoh K, Sawano J (1991) Male germ-line chromosomes of *Orthetrum poecilops miyajimaensis* Yuki et Doi, 1938 (Libellulidae: Odonata). *Tombo* 34: 29–30.
- Toyoshima H, Hirai H (1953) Studies on chromosomes of four dragonflies from Kagawa Prefecture. *Kagawa Biology* 1: 17–19. [In Japanese]
- Thomas KI, Prasad R (1981) The chromosomes of five Indian dragonflies (Odonata). *Perspectives in Cytology and Genetics* 3: 629–632.
- Thomas KI, Prasad R (1986) A study of the germinal chromosomes and C-band patterns in four Indian dragonflies (Odonata). *Perspectives in Cytology and Genetics* 5: 125–131.
- Tyagi BK (1977) A note on the karyotypes of *Burmagomphus pyramidalis* Laidlow and *Onychogomphus saundersi duaricus* Faser (Anizoptera; Gomphidae). *Odonatologica* 6(4): 277–282.
- Tyagi BK (1978a) The chromosome numbers and sex-determining mechanisms newly recorded in thirteen Indian dragonflies (Odonata). *Chromosome Information Service*, Tokyo 25: 5–7.



- Tyagi BK (1978b) Studies on the chromosomes of Odonata of Dun Valley (Dehradun, India). PhD thesis, University of Garhwal, Srinagar.
- Tyagi BK (1982) Cytotaxonomy of Indian dragonflies. *Indian Review of Life Sciences* 2: 149–161.
- van Brink JM, Kiauta B (1964) Notes on chromosome behaviour in the spermatogenesis of the damselfly *Enallagma cyathigerum* (Charp.) (Odonata: Coenagrionidae). *Genetica* 35: 171–174. <https://doi.org/10.1007/BF01804885>
- Walia GK (2007) Cytomorphological studies on *Gynacantha milliardi* Fraser of the family Aeschnidae (Anisoptera: Odonata). *Cytologia* 72(1): 57–62. <https://doi.org/10.1508/cytologia.72.57>
- Walia GK, Chahal SS (2014) Distribution of constitutive heterochromatin and nucleolar organizer regions in two species of family Gomphidae (Odonata: Anisoptera). *Nucleus* 57: 223–227. <https://doi.org/10.1007/s13237-014-0122-z>
- Walia GK, Chahal SS (2018) Cytogenetic characterization of *Macromia moorei* Selys, 1874 of family Macromiidae (Odonata: Anisoptera) from India by C-banding, silver nitrate staining and sequence specific staining. *International Journal of Life Sciences Research* 6(2): 64–68.
- Walia GK, Chahal SS (2019) Cytogenetic report on *Cordulegaster brevistigma* and *Watanabeopetalia atkinsoni* (Odonata: Cordulegastridae, Chlorogomphidae). *Odonatologica* 48(1–2): 101–113.
- Walia GK, Chahal SS (2020) Linear differentiation of chromosomes of *Anisogomphus bivittatus* Selys, 1854 from India (Odonata: Anisoptera: Gomphidae). *International Journal of Entomology* 5(2): 120–122.
- Walia GK, Chahal SS, Babu R (2016) Cytogenetic report on *Gynacanthaeschna sikkima* from India (Odonata: Aeshnidae). *Odonatologica* 45: 87–94.
- Walia GK, Chahal SS, Somal DS (2018) Chromosome observations based on C-banding, Ag-NOR and sequence-specific staining in two *Anax* species from India (Odonata: Aeshnidae). *Odonatologica* 47(1–2) 2018: 145–160.
- Walia GK, Devi M (2018) Distribution of constitutive heterochromatin in four species of genus *Copera* of family Platycnemididae (Odonata: Zygoptera) from India. *International Journal of Life Sciences* 6(2): 457–461.
- Walia GK, Devi M (2020a) Cytogenetic characterization of five species of genus *Coeliccia* of family Platycnemididae (Odonata: Zygoptera) using C-banding, silver nitrate staining and sequence specific staining. *Nucleus* (2020). <https://doi.org/10.1007/s13237-020-00314-3>
- Walia GK, Devi M (2020b) Cytogenetic data of subfamily Disparoneurinae (Odonata: Zygoptera: Platycnemididae) based on localization of C-heterochromatin, AgNOR's and AT-GC regions. *International Journal of Entomology Research* 5(2): 70–73.
- Walia GK, Katnoria N (2018) Morphological variation in the chromosome complement of *Neurobasis chinensis chinensis* of family Calopterygidae (Odonata: Zygoptera). *International Journal of Life Sciences Research* 6(4): 260–266.
- Walia GK, Katnoria N, Gill JK (2018) Chromosomes of *Libellago lineata lineata* (Chlorocyphidae: Odonata). *Indian Journal of Entomology* 80(3): 737–740. <https://doi.org/10.5958/0974-8172.2018.00118.9>
- Walia GK, Kaur H, Kaur J (2011) Karyotypic variations in the chromosome complement of *Pantala flavescens* (Fabricius) of the family Libellulidae (Anisoptera: Odonata). *Cytologia* 76(3): 301–307. <https://doi.org/10.1508/cytologia.76.301>



- Walia GK, Kaur H, Kaur J (2015) Karyomorphological variations in the chromosome complement of *Orthetrum taeniolatum* of family Libellulidae (Odonata: Anisoptera). *Cytologia* 80(1): 95–99. <https://doi.org/10.1508/cytologia.80.95>
- Walia GK, Sandhu R (1999) Karyotypic study of two species of family Aeschnidae (Anisoptera: Odonata). *Chromosome Science* 3: 45–47.
- Walia GK, Sandhu R (2002) Chromosomal data on seven species of genus *Orthetrum* (Libellulidae: Anisoptera: Odonata). *Bionature* 22: 7–12.
- Walia GK, Sandhu R, Goyal S (2006) Cytogenetical analysis of *Nepogomphus modestus* from Palampur area of Himachal Pradesh, India (Gomphidae: Anisoptera). *Chromosome Science* 9(3): 99–100.
- Wasscher M (1985) The karyotypes of some dragonflies from Kenya and Sudan. *Notulae odonatologicae* 2(6): 105–106.
- Wolfe LS (1953) A study of the genus *Uropetala* Selys (order Odonata) from New Zealand. *Transactions and Proceedings of the Royal Society of New Zealand* 80(3–4): 245–275.
- Zhu H, Wu J (1986) Notes on the male germ cell karyotypes of some Odonata from the Shanxi Province, China. *Notulae odonatologicae* 2: 118–120.